Effects of pomegranate juice consumption on inflammatory markers in patients with type 2 diabetes: A randomized, placebo-controlled trial

Golbon Sohrab, Javad Nasrollahzadeh, Hamid Zand¹, Zohreh Amiri¹, Maryam Tohidi², Masoud Kimiagar

Department of Clinical Nutrition and Dietetics, Faculty of Nutrition Sciences and Food Technology, ¹Department of Basic Sciences and Cellular and Molecular Nutrition, Faculty of Nutrition Sciences and Food Technology, ²Prevention of Metabolic Disorders Research Center, Research Institute for Endocrine Sciences, Shahid Beheshti University of Medical Sciences, Tehran, Iran

Background: Diabetes causes the increased concentration of circulatory cytokines as a result of inflammation. Considering that pomegranate juice (PJ) is known to have antioxidant and anti-inflammatory properties, the purpose of this study was to determine the effects of PJ consumption on markers of inflammation in patients with type 2 diabetes (T2D). **Materials and Methods:** In a randomized, double-blind clinical trial study, 50 patients with T2D (40-65 years old) were randomly assigned to one of two groups. Participants in each group received either 250 mL/day PJ or a control beverage for 12 weeks. Biochemical markers including fasting plasma glucose (FPG), insulin and inflammatory markers were assayed on the baseline and follow-up blood samples. **Results:** In all, 44 patients in two groups were included in the analysis: PJ (n = 22) and placebo (n = 22). After 12 weeks of intervention, in the PJ group, there were 32% and 30% significant decreases in plasma C-reactive protein (hs-CRP) and Interlukin-6, respectively (P < 0.05). The mean \pm SD plasma interlukin-6 (7.1 ± 5.6 vs. 11.9 ± 14.4 mg/L) and hs-CRP (1791 ± 1657 and 1953 ± 1561 ng/mL) concentrations in the PJ group were significantly lower than the placebo group after intervention (P < 0.05). **Conclusion:** PJ consumption by patients with T2D does not affect FPG or the insulin resistance index (HOMA-IR), whereas it does reduce Interlukin-6 and hs-CRP concentrations in plasma. Therefore, PJ consumption may have an anti-inflammatory effect in patients with T2D.

Key words: Diabetes mellitus, inflammation, pomegranate

How to cite this article: Sohrab G, Nasrollahzadeh J, Zand H, Amiri Z, Tohidi M, Kimiagar M. Effects of pomegranate juice consumption on inflammatory markers in patients with type 2 diabetes: A randomized, placebo-controlled trial. J Res Med Sci 2014;19:215-20.

INTRODUCTION

Type 2 diabetes (T2D) is one of the main noncommunicable chronic diseases, and its complications have become a major cause of morbidity and mortality worldwide. The increase in adipose tissue mass observed in obesity can lead to chronic activation of the innate immune system, which can lead to insulin resistance and T2D.^[1] All of the following, heart disease, metabolic syndrome and T2D, have in common the increased concentration of circulatory cytokines as a result of inflammation.^[2] Low-grade systemic inflammation is characterized by a two- to threefold increase in systemic plasma concentrations of cytokines such as tumor necrosis factor (TNF)-α, interleukin (IL)-6 and C-reactive protein (CRP).^[1] The cytokines produced by adipose tissue have a key role in promoting atherosclerosis and, therefore, cardiovascular disease (CVD).^[2]

Pomegranate juice (PJ) contains diverse groups of polyphenols, including ellagitanins, gallotannins, and ellegic acid, as well as flavonoids, such as anthocyanins.^[3] However, its antioxidant activity is mainly due to hydrolysable tannin, including punicalagins, anthocyanins and ellagic acid.^[4] PJ antioxidant activity was found to be three times higher than that of red wines and green teas, and two- to sixfolds stronger than the other natural beverages.^[5]

Evidence for the clinical benefits of PJ has been reported in some studies,^[6-8] but few studies that have assessed its impact on inflammation have been conducted *in vitro* or used pomegranate extract.^[9,10] Although patients with T2D are exposed to severe systemic oxidative stress and inflammation, data on the effect of PJ intake among these patients is limited. In this study, the effects of PJ consumption on markers of inflammation in patients with T2D were investigated.

MATERIALS AND METHODS

This study was a randomized, double-blind, placebocontrolled trial.

Address for correspondence: Dr. Maryam Tohidi, Prevention of Metabolic Disorders Research Center, Research Institute for Endocrine Sciences, Shahid Beheshti University of Medical Sciences, Tehran, Iran. E-mail: Tohidi@endocrine.ac.ir Received: 01-09-2013; Revised: 25-11-2013; Accepted: 08-01-2014

Subjects and ethical aspects

T2D patients, aged 40-65 years, with a diagnosis of at least 1 year, were recruited from the Charity Foundation for Special Diseases and Health Center of District 2 of Tehran, Iran. All the patients controlled their diabetes with oral hypoglycemic agents. Patients were excluded from the trial if they were smoking, suffering from any other chronic diseases and taking estrogen or progesterone (if female) or antioxidant supplements or used insulin as diabetes medication.

At baseline, patients were stratified by sex and randomly assigned to one of two groups: Group A (PJ, n = 25) and group B (placebo, n = 25). Random allocation of patients to treatment groups was performed by sequentially numbered containers. Randomization was performed by an assistant and the group allocation was blinded for the investigator and participants.

Written informed consent was obtained from all patients. Ethic approval for the trial was obtained from the ethical committee of the National Nutrition and Food Technology Research Institute (Tehran, Iran). This clinical trial has been registered in the Iranian Registry of Clinical Trials at http://www.irct.ir with the following identification: IRCT201206144010N8, and was conducted between November 2012 and March 2013.

Protocol

At baseline, patients were stratified by sex and randomly assigned to consume 250 mL/day PJ or a control beverage of similar color and energy content for 12 weeks. The study product was packaged in single-serving bottles labeled such that neither subjects nor staff members were aware of the treatment assignment.

Subjects were advised not to change their dietary habits, physical activities or drug medication. The dietary intakes of subjects were assessed using a 3-day dietary recall (two weekdays and one weekend day) at baseline and at the end of 12 weeks. Patients' diets were analyzed by the Nutritionist IV software (N Squared Computing, San Bruno, CA, USA).

Measurements

At baseline and after 12 weeks of intervention, 10 mL blood was collected from each patient after a 12-14-h overnight fasting. Blood samples to which an anti-coagulating agent was added were centrifuged at 4000 rpm for 10 min. The plasma samples were separated into aliquots and were frozen at -70°C until they were assayed. Fasting plasma glucose (FPG) concentration was assessed using the enzymatic colorimetric method using glucose oxidase by commercial kits (Pars Azemoon, Tehran, Iran) and a Selectra 2 auto-analyzer (Vital Scientific, Spankeren, the Netherlands). The coefficient of variation (CV) for FPG was 1.3%. Plasma insulin and inflammatory markers were assessed using the enzyme-linked immunosorbent assay method (ELISA). Insulin was measured using a commercial kit (Mercodia, Uppsala, Sweden), with a CV of 5.0%. The plasma concentration of TNF- α and high-sensitive CRP (hs-CRP) were assayed by related kits (Komobiotech Inc., Gangseo-gu Seoul, Korea) and Diagnostics (Biochem, Canada Inc., Ontario, Canada), respectively. Interlukin-6 was measured using commercial kits (Biolegend, CA, USA). All immunoassays were performed by an ELISA reader Sunrise, Tecan Co. Salzburg, Austria. CVs for TNF- α , hs-CRP and IL-6 were 6.7, 6.3 and 7.1, respectively.

Weight and height were measured using a balance Seca scale with a stadiometer at the baseline and at the end of the intervention. Weight was measured while the subjects were minimally clothed and not wearing shoes, and was recorded to the nearest 100 g. Height was measured while subjects were standing without shoes, with their shoulders in a normal position, and was recorded to the nearest 0.5 cm. Body mass index (BMI) was calculated as weight (kg) divided by square of height (m).

In addition, HOMA-IR was calculated as: [Glucose (mmol/L) \times Insulin (μ U/mL)]/22.5.

Compliance

To ascertain patient compliance, we provided each patient with a fixed number of PJ bottles and instructions to return the unused bottles at the end of the study. Based on the number of returned bottles by each patient, their compliance was determined, which was 90% for our participants. No adverse events were reported.

Pomegranate and placebo juice

In order to choose the commercial PJ with the highest polyphenol levels, several hand-squeezed and various commercially available juices were analyzed using the colorimetric assay. The phenols were determined by the Folin-Ciocalteau reagent, using Gallic acid as a standard.^[11] Total flavonoid content was measured by the aluminum chloride colorimetric assay using Catechin as a standard.^[12] PJ was diluted 1:10 (v:v) to measure its total antioxidant capacity (TAC), which was based on the inhibition percent of ABTS and comparison with bovine serum albumin (BSA) standard curve.^[13] PJ composition is shown in Table 1.

PJ and placebo was prepared by Alifard Inc., Tehran, Iran. We used special essence and formula of Wonderful Variety, Pom, supplied by Roll International Corporation, Los Angeles, CA, USA, for preparing the placebo. The sugar content of the placebo was 50% of glucose and 50% of fructose. Analyzing the placebo juice using the colorimetric assay verified that it had no polyphenols. The juice and the placebo were kept at room temperature (<25°C) until opened, as recommended by the manufacturer.

Sample size estimation and statistical analysis

The results are expressed as mean \pm SD for quantitative variables and n (%) for qualitative variables, and differences were considered significant at *P* \leq 0.05.

Statistical analysis of data was performed using the Statistical Package for the Social Sciences (SPSS Inc., Chicago, IL, USA) for Windows version 16.0. A χ^2 test was used to compare the qualitative variables between the two groups.

Because all quantitative parameters according to the Kolmogorov–Smirnov test had normal distributions, a paired *t*-test was used to compare the pre- and postintervention variables within groups, and the mean changes within groups were tested using the Student's *t*-test. Adjustment for differences in baselines covariates and changes in variables during the study were performed by analysis of covariance using general linear models.

RESULTS

The sample size was designed to detect a 2 ng/L difference of plasma IL-6 between the groups with 95% confidence interval and 90% power.^[14] In all, 44 patients [PJ (n = 22), placebo (n = 22)] were included in the analysis as presented in the flow chart [Figure 1].

The baseline characteristics of patients did not differ significantly between the PJ-treated and the placebo groups [Table 2]. Anthropometric factors had no significant differences between the two groups at the baseline or at the end of Week 12; in addition, these factors did not change significantly within the groups during the study [Table 3].

Dietary energy and carbohydrate intake showed significant differences between the two groups at the baseline and at the end of the study (P < 0.05); however, none of the dietary factors changed significantly within each group during the study [Table 3].

No serious adverse events or side-effects were reported. All analyses were adjusted for energy and carbohydrate intake by analysis of covariance.

Plasma interlukin-6 and hs-CRP concentration reduced significantly in the PJ group at the end of the 12^{th} week compared with baseline (P < 0.05), whereas no significant change was observed in the placebo group. Decreases in plasma interlukin-6 and hs-CRP concentrations in the PJ

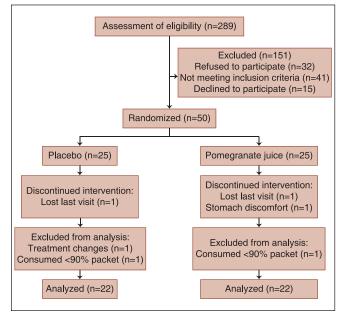


Figure 1: Flow chart of the study

Table 1: Composition of pomegranate juice			
	Pomegranate juice	Placebo	
Total polyphenols (mg/L)	1946	0	
Total flavonoid (µg/mL)	345.87	0	
TAC (mmol/L) BSA	7	0	
Energy (kcal)*	126	126	
Sugar (g)*	24	24	
Protein (g)*	1.4	0	
Fiber (g)*	0	0	

Values are per 250 mL; TAC = Total antioxidant capacity; BSA = Bovine serum albumin

Table 2: Baseline characterist	ics of patients in t	the
pomegranate juice-treated and	d placebo groups	
		DI 1

Characteristics	Pomegranate	Placebo	
	(<i>n</i> = 22)	(<i>n</i> = 22)	
Age (years) ¹	55±6.7	56.9±6.8	
Duration of diabetes (years) ¹	5.8±3.2	6.5±4.3	
Sex			
Men	11 (50%)	12 (54.5%)	
Women	11 (50%)	10 (45.5%)	
Anti-hypertensive drugs (%)	4 (18.2%)	6 (27.3%)	
(ACE-I or ARB)	3 (13.6%)	4 (18.2%)	
Anti-hyperlipidemic drugs (%)			

¹Age and duration of diabetes are presented as mean \pm SD; ACEI = Angiotensin converting enzyme inhibitors; ARB = Angiotensin receptor blockers; *P* - value > 0.05

group were significant in comparison with the placebo group (P < 0.05; Table 4), plasma TNF- α concentration did not change significantly within each group or between groups during the study [Table 4].

Mean differences of inflammatory variables compared with baseline values are presented in Table 4. There were 30% and 32% significant decreases in plasma IL-6 and hs-CRP in the PJ and placebo groups (P < 0.05), respectively.

pomegranate juice-treated and placebo groups ¹			
Factors	Groups	Baseline	Week 12
Weight (kg)	Pomegranate	76.9±15	77.4±15.4
	Placebo	75.2±15	74.9±9.7
BMI (kg/m²)	Pomegranate	29.4±3.9	29.4±3.9
	Placebo	28.6±4.2	28.4±4.4
Energy (kcal/d)	Pomegranate	1784±238 ^b	1787±233 ^b
	Placebo	2028±241	2011±221
Protein (g/d)	Pomegranate	54±12	58±12
	Placebo	59±10	65±9
Carbohydrate (g/d)	Pomegranate	234±35 ^b	266±41 ^b
	Placebo	232±27	257±37
Fiber (g/d)	Pomegranate	15±4	15±3
	Placebo	16±3	15±4
Fat (g/d)	Pomegranate	73±20	72±19
	Placebo	84±15	83±19
SAFA (g/d)	Pomegranate	16.2±5.5	17.2±6.4
	Placebo	23.7±8.8	21.3±8.2
MUFA (g/d)	Pomegranate	30.7±11	29.8±8.8
	Placebo	32.5±6.1	33.6±8.7
PUFA (g/d)	Pomegranate	21.4±6.7	19.4±4.4
	Placebo	20.5±6.1	21.7±5.9
Cholesterol (mg/d)	Pomegranate	134.3±60.7	136.1±48.6
	Placebo	169.6±78	189.9±58.2
Vitamin E (mg/d)	Pomegranate	22.7±5.9	20.7±4.7
	Placebo	20.5±6.6	22.1±6.6
Vitamin C (mg/d)	Pomegranate	78.7±54.3	77.3±32.9
/	Placebo	60.1±32.8	66.6±32.1
Vitamin A (UI/d)	Pomegranate	559.7±478.3	719.9±609
/	Placebo	440.8±349.4	359.7±170.2

 Table 3: Anthropometric and dietary factors in the

 pomegranate juice-treated and placebo groups¹

All values are presented as mean \pm SD; $^{1}n = 22$ for all values; BMI = Body mass index; MUFA = Monounsaturated fatty acids; SAFA = Saturated fatty acids; PUFA = Polyunsaturated fatty acids; a) P < 0.05 vs baseline, b) P < 0.05 vs the placebo group

PJ consumption by the patients did not significantly affect the levels of parameters such as FPG, insulin and HOMA-IR [Table 4].

DISCUSSION

We found that consumption of PJ had beneficial effects on some inflammatory markers in patients with T2D through a significant decrease in IL-6 and hs-CRP in plasma. Adjustment for confounding factors including energy and carbohydrate intake provide the same results.

Although there are some clinical trials that have described the effects of PJ consumption on the oxidative state and blood pressure,^[6,15-17] only a limited number of *in vitro* studies have assessed the impact of pomegranate on inflammation using the pomegranate extract.^[9,10] To the best of our knowledge, this is the first study that examines the effects of PJ consumption on markers of inflammation in individuals with T2D.

A 1-year intake of PJ by hemodialysis patients showed a significant reduction in the level of all oxidative stress and inflammatory biomarkers compared with placebo, with a significant reduction in IL6 and TNF- α .^[3] Decreased circulating levels of vascular cell adhesion molecule 1 (VCAM-1) were reported with the consumption of PJ for a period of 2 weeks in hypertensive subjects, but no significant effect was observed from PJ on the serum levels of intercellular adhesion molecule 1 (ICAM-1), hs-CRP, IL-6 and lipid profile.^[18]

Table 4: Plasma concentrations of fasting glucose, insulin, HOMA-IR, inflammatory markers in the pomegranate	÷
juice-treated and placebo groups ¹	

Parameters	Groups	Baseline	Week 12	P-value (within groups)	Mean changes ²	Mean changes ³ (%)
Fasting plasma glucose (mg/dL)	Pomegranate	160.3±47.8	162.3±35.2	NS	2±45.3	6±27
	Placebo	148.7±42.1	158.6±49.1	NS	9.8±43.3	10±35
P-value (between groups)		NS	NS		NS	NS
Insulin (mU/L)	Pomegranate	7.6±3.8	7.1±2.6	NS	-0.52±3.2	-0.45±28
	Placebo	7.6±3.8	7.1±3.1	NS	-0.48±2.4	-3±27
P-value (between groups)		NS	NS		NS	NS
HOMA-IR	Pomegranate	54.3±29.9	51.4±21.8	NS	-2.9±29.7	6±40
	Placebo	48.4±20.3	47.0±16.7	NS	-1.3 ± 15.9	4±34
P-value (between groups)		NS	NS		NS	NS
hs-CRP (ng/mL)	Pomegranate	3243±2935	1791±1657	<i>P</i> <0.01	-1441±2251	-32±31.8
	Placebo	2667±1577	1953±1561	<i>P</i> <0.05	-271±1322	-2±41.7
P-value (between groups)		NS	<i>P</i> <0.05		<i>P</i> <0.05	<i>P</i> <0.05
Interlukin-6 (ng/L)	Pomegranate	10.9±4.4	7.1±5.6	<i>P</i> <0.01	-3.8±6.1	-30±46
	Placebo	12.5±14.3	11.9±14.4	NS	-0.6±2.6	-5±25
P-value (between groups)		NS	<i>P</i> <0.05		<i>P</i> <0.05	<i>P</i> <0.05
TNF- α (ng/L)	Pomegranate	37±19.3	30.4±17.5	<i>P</i> <0.01	-6.6±9.2	-16 ± 17
	Placebo	36±20.4	30.8±18.0	<i>P</i> <0.01	-5.2±7.1	-11±22
<i>P</i> -value (between groups)		NS	NS		NS	NS

All values are presented as mean ± SD; ¹n = 22 for all values; ²Mean changes reflect Week 12 – baseline values; ³Mean changes% reflect (mean changes/baseline)*100; NS = None significant; HOMA-IR = Homeostatic model assessment for Insulin Resistance; Hs-CRP = High sensitive C-reactive protein

In vitro studies have shown the positive inhibitory effect of pomegranate punicic acid on the production of TNF-α, induced by priming of reactive oxygen species,^[19] on the modulation of inflammatory cell signaling in colon cancer cells via pomegranate ellagitannins^[9] and reducing pro-inflammatory cytokine production via inhibiting the gene expression (pomegranate fruit extract).^[10] However, findings from *in vitro* studies often conflict with findings obtained from *in vivo* ones.

Most studies investigating the antioxidant and anti-inflammatory effects of polyphenols and flavonoids have been conducted *in vitro*. It has been indicated that flavonoid metabolites have different biological and antioxidant properties than their parent compounds, which suggested that data from *in vitro* studies using nonmetabolites of flavonoids are of limited relevance to *in vivo* studies.^[20]

Low-grade systemic inflammation such as diabetes is characterized by an increase in systemic plasma concentrations of cytokines such as TNF- α , IL-6 and CRP.^[1] In our study, IL-6 and hs-CRP decreased significantly but TNF- α did not show any significant reduction.

Zhao *et al.* have suggested that plasma TNF- α level may not be detectable in a single blood sampling as this may not reflect the host production of this inflammatory mediator.^[21] Human blood mononuclear cells are known to synthesize and secrete TNF- α . It has been suggested that activated monocytes and macrophages are important cellular sources for circulating pro-inflammatory cytokines.^[22]

The main antioxidant compounds in PJ are hydrolysable tannins, anthocyanins and ellagic acid derivatives, which contribute to the total antioxidant capacity of the juice.^[4] Some mechanisms have been also proposed for the anti-inflammatory properties of PJ, including inhibition of enzymes related to inflammation, such as peroxisome proliferators active receptors (PPARs), nuclear transcription factor kappa B (NF-kB), and NSAID activated gene-1 (NAG-1),^[23] scavenging-free radicals^[4] or production of urate, which is stimulated by fructose and other compositions of flavonoid-rich foods.^[24] In addition, pomegranate enhances endothelial nitric oxide production and bioavailability^[25] and protects nitric oxide against oxidative destruction.^[26]

Strengths of the current study are a randomized, doubleblind, placebo-controlled trial design, acceptable response rate and detailed data collection through face-to-face meetings. Moreover, PJ and placebo were provided for all patients. This study had limitations because we could not measure plasma polyphenol derivatives' levels nor did we evaluate the level of TNF- α in the blood mononuclear cells. Hence, it is suggested these two be measured in future studies.

In conclusion, our findings suggest that PJ, which is a source of natural sugars, does not affect FPG and insulin resistance index (HOMA-IR) in patients with T2D, and it also acts as an anti-inflammatory agent, lowering some inflammatory factors including IL-6 and hs-CRP. Further studies with a longer intervention period as well as a bigger sample size or using PJ with different polyphenol contents are needed in order to evaluate its protective role against diabetes and its complications and the antiinflammatory effects of PJ.

ACKNOWLEDGMENTS

This study was supported by the National Nutrition and Food Technology Research Institute of Iran and The Research Institute for Endocrine Sciences with research project number of 435. The authors would like to express their gratitude to the subjects for their participation and cooperation in this research and Alifard Inc., Tehran, Iran for preparing the pomegranate and placebo juices. Special thanks are due to Wonderful Variety, Pom; supplied by Roll International Corporation, Los Angeles, CA, USA for their placebo formulation. The authors wish to acknowledge Ms. Niloofar Shiva for her critical editing of English grammar and syntax of the manuscript.

REFERENCES

- 1. Calle MC, Fernandez ML. Inflammation and type 2 diabetes. Diabetes Metab 2012;38:183-91.
- 2. Puglisi M, Fernandez ML. Modulation of C-reactive protein, tumor necrosis factor-alpha and adiponectin by diet, exercise and weight loss. J Nutr 2008;138:2293-6.
- Shema-Didi L, Sela S, Ore L, Shapiro G, Geron R, Moshe G, et al. One year of pomegranate juice intake decreases oxidative stress, inflammation, and incidence of infections in hemodialysis patients: A randomized placebo-controlled trial. Free Radic Biol Med 2012;53:297-304.
- 4. Gil MI, Tomas-Barberan FA, Hess-Pierce B, Holcroft DM, Kader AA. Antioxidant Activity of Pomegranate Juice and Its Relationship with Phenolic Composition and Processing. J Agric Food Chem 2000;48:4581-9.
- 5. Seeram NP, Aviram M, Zhang Y, Henning SM, Feng L, Dreher M, et al. Comparison of antioxidant potency of commonly consumed Polyphenol-Rich beverages in the United States. J Agric Food Chem 2008;56:1415-22.
- 6. Aviram M, Rosenblat M, Gaitini D, Nitecki S, Hoffman A, Dornfeld L, *et al*. Pomegranate juice consumption for 3 years by patients with carotid artery stenosis reduces common carotidintimamedia thickness, blood pressure and LDL oxidation. Clin Nutr 2004;23:423-33.
- 7. Rosenblat M, Hayek T, Aviram M. Anti-oxidative effects of pomegranate juice (PJ) consumption by diabetic patients on serum and on macrophages. Atherosclerosis 2006;187:363-71.
- 8. Esmaillzadeh A, Tahbaz F, Gaieni I, Alavi-Majd H, Azadbakht L. Concentrated pomegranate juice improves lipid profiles in diabetic patients with hyperlipidemia. J Med Food 2004;7:305-8.

- Adams LS, Seeram NP, Aggarwal BB, Takada Y, Sand D, Heber D. Pomegranate juice, total pomegranate ellagitannins, and punicalagin suppress inflammatory cell signaling in colon cancer cells. J Agric Food Chem 2006;54:980-5.
- Rasheed Z, Akhtar N, Anbazhagan AN, Ramamurthy S, Shukla M, Haqqi TM. Polyphenol-rich pomegranate fruit extract (POMx) suppresses PMACI-induced expression of pro-inflammatory cytokines by inhibiting the activation of MAP Kinases and NFkappaB in human KU812 cells. J Inflamm (Lond) 2009;6:1.
- 11. Bray HG, Thorpe WV. Analysis of phenolic compounds of interest in metabolism. Methods Biochem Anal 1954;1:27-52.
- 12. Hajimahmoodi M, Oveisi MR, Sadeghi N, Jannat B, Hadjibabaie M, Farahani E, *et al.* Antioxidant properties of peel and pulp hydro extract in ten Persian pomegranate cultivars. Pak J Biol Sci 2008;11:1600-4.
- Cano A, Herna'ndez-Ruiz J, Garcı'a-Ca'novas F, Acosta M, Arnao MB. An end-point method for estimation of the total antioxidant activity in plant material. Phytochem Anal 1998;9:196-202.
- 14. Noori N, Tabibi H, Hosseinpanah F, Hedayati M, Nafar M. Effects of combined administration of Lipoic Acid and Pyridoxine on serum systemic and vascular inflammatory factors in patients with diabetic nephropathy. Iran J Endocrinol Metab 2010;12:99-110.
- 15. Rosenblat M, Hayek T, Aviram M. Anti-oxidative effects of pomegranate juice (PJ) consumption by diabetic patients on serum and on macrophages. Atherosclerosis 2006;187:363-71.
- Rock W, Rosenblat M, Miller-Lotan R, Levy AP, Elias M, Aviram M. Consumption of wonderful variety pomegranate juice and extract by diabetic patients increases paraoxonase 1 association with highdensity lipoprotein and stimulates its catalytic activities. J Agric Food Chem 2008;56:8704-13.
- 17. Parsaeyan N, Mozaffari-Khosravi H, Mozayan MR. Effect of pomegranate juice on paraoxonase enzyme activity in patients with type 2 diabetes. J Diabetes Metab Disord 2012;11:11.

- Asgary S, Sahebkar A, Afshani MR, Keshvari M, Haghjooy Javanmard S, Rafieian-Kopaei M. Clinical evaluation of blood pressure lowering endothelial function improving, hypolipidemic and anti-inflammatory effects of Pomegranatejuice in hypertensive subjects. Phytother Res 2013 Mar 21 [In Press].
- 19. Boussetta T, Raad H, Lettéron P, Gougerot-Pocidalo MA, Marie JC, Driss F, *et al.* Punicic acid a conjugated linolenic acid inhibits TNF alpha-induced neutrophil hyperactivation and protects from experimental colon inflammation in rats. PLoS One 2009;4:e6458.
- Lotito SB, Zhang WJ, Yang CS, Crozier A, Frei B. Metabolic conversion of dietary flavonoids alters their anti-inflammatory and antioxidant properties. Free Radic Biol Med 2011;51:454-63.
- Zhao SP, Xu TD. Elevated tumor necrosis factor alpha of blood mononuclear cells in patients with congestive heart failure. Int J Cardiol 1999;71:257-61.
- 22. Tracy K, Cerami A. Tumor necrosis factor, other cytokines and disease. Annu Rev Cell Biol 1993;9:317-43.
- 23. Yoon JH, Baek SJ. Molecular targets of dietary polyphenols with anti-inflammatory properties. Yonsei Med J 2005;46:585-96.
- 24. Lotito S, Frei B. Consumption of flavonoid-rich food and increased plasma antioxidant capacity in humans: Cause, consequence, or epiphenomenon? Free Radic Biol Med 2006;41:1727-46.
- 25. Schmitt CA, Dirsch VM. Modulation of endothelial nitric oxide by plant-derived products. Nitric Oxide 2009;21:77-91.
- Ignarro L, Byrns R, Sumi D, Nigris F, Napoli C. Pomegranate juice protects nitric oxide against oxidative destruction and enhances the biological actions of nitricoxide. Nitric Oxide 2006;15:93-102.

Source of Support: This study was supported by the National Nutrition and Food Technology Research Institute of Iran and the Research Institute for Endocrine Sciences, **Conflict of Interest:** None declared.