Associations between high density lipoprotein mean particle size and serum paraoxonase-1 activity

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Background: High density lipoprotein (HDL) particles are heterogeneous in composition, structure, size, and may differ in conferring protection against cardiovascular disease. HDL associated enzyme, paraoxonase-1 (PON1), has an important role in attenuation of atherogenic low density lipoprotein (LDL) oxidation. The aim of this study was to investigate the associations between HDL particle size and PON1 activity in relation to serum HDL cholesterol (HDL-C) levels. **Materials And Methods:** One hundred and forty healthy subjects contributed to this study. HDL was separated by sequential ultracentrifugation and its size was estimated by dynamic light scattering. Paraoxonase activity was measured spectrophotometrically using paraoxon as substrate. **Results:** Results of this study showed that PON1 activity had negative correlations with HDL mean particle size (r = -0.22, P < 01), HDL₂/HDL₃ ratio, and serum HDL-C levels (r = -0.25, P < 0.01). HDL mean particle size and HDL₂/HDL₃ ratio had negative correlation with body mass index (BMI), waist hip ratio (WHR), and serum triglyceride (TG) levels, and positive correlation with serum HDL-C levels. Serum HDL-C levels had significant positive correlations with age, total cholesterol (TC), and apolipoprotein A-I (apo A-I) and significant negative correlation with BMI, WHR, and TG. **Conclusion:** Based on the results of this study, determination of HDL mean particle size beside the serum PON1 activity may help to better understand the CAD risks, pathogenesis, and prognosis, and may also help to design therapeutic protocols toward beneficial modifications of HDL characteristics.

Key words: Dynamic light scattering, high density lipoprotein size, HDL-C, paraoxonase-1 activity, zetasizer

INTRODUCTION

There is a strong inverse relationship between plasma high density lipoprotein (HDL) level and the risk of developing coronary artery disease (CAD). [1,2] The major role of HDL is the reverse cholesterol transport, which leads to cholesterol movement from peripheral tissue and vessel wall to the liver. [3] Anti-inflammatory, antioxidant, anticoagulant, and profibrinolytic actions of HDL are among other functions that may contribute further to its ability to protect against CAD. [3] HDL particles are heterogeneous in their composition, structure, and size. Several methods such as immunoaffinity chromatography, ultracentrifugation. [4] gradient gel electrophoresis. [5] electron microscopy. [6] and nuclear magnetic resonance (NMR) have been used to isolate and characterize HDL subfractions and at least five

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subfractions of HDL with sizes of 7.3-13 nm have been identified.[7] One of the best known anti-inflammatory and antioxidant functions of HDL is its ability to inhibit the oxidation of low-density lipoprotein (LDL).[8] HDL exerts its inhibitory effects on oxidative modification of LDL, in part, by its related enzyme paraoxonase-1 (PON1) which can hydrolyze lactones, and several non-physiological substrates, such as aryl esters and organophosphates.^[9] PON1 is transported in plasma as a component of HDL, and many studies have also shown that PON1 inhibits LDL oxidation in vitro.[10] There is a close physiological association between PON1 and HDL in plasma. HDL facilitates the secretion of the PON1 by the liver, stabilizes the enzyme,[11] and provides a hydrophobic environment which is needed for PON1 function.[12] In return, PON1 prevents the oxidation of HDL.[13] There is a wide inter-individual variation in PON1 activity and concentrations due to variation in PON1 gene, and also, because of the influences of lifestyle factors such as smoking, alcohol consumption, etc. [14,15] A study of populations with high prevalence of atherosclerosis suggested that PON1 could influence HDL concentrations in patients with familial hypercholesterolemia.[16] It has been demonstrated that PON1 is present in small dense subclasses of HDL particles including apolipoprotein A-I (apo A-I) and

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apolipoprotein J.^[17] Investigations have also indicated that there is a relationship between PON1 activity and HDL size with CAD risk.^[18,19] However, this relationship is not demonstrated in all studies.^[20,21]

With these in mind, the protective role of HDL against CAD seems to be a complicated and multi-factorial procedure that depends on the HDL particle size, number, composition, distribution, and its related enzymes such as PON1. The aim of this study was to investigate the associations between HDL mean particle size and serum PON1 activity in relation to serum HDL-C levels in healthy subjects.

MATERIALS AND METHODS

Subjects

For this study, 140 healthy men between 20 and 50 years of age from Isfahan area were recruited to Isfahan Cardiovascular Research Center between June and September 2010. All subjects were healthy, non-smoking, non-obese [body mass index (BMI) ≤30], and non-diabetic [fasting blood sugar (FBS) ≤110 mg/dl] volunteers, without familiar hypercholesterolemia and any sign of cardiovascular or inflammatory diseases, and no one was taking lipid-modifying medication. All participants were interviewed and a checklist relating to their age, medical history, and lifestyle information was completed. All participants signed an informed consent document approved by the Isfahan University of Medical Sciences Medical Ethics Committee. This study was funded by grant number 188140 from Isfahan University of Medical Sciences, Isfahan, Iran.

Anthropometric measurements

Weight was measured while subjects were minimally clothed without shoes and height was measured in a standing position. BMI was calculated as weight in kilograms divided by height in meters squared. Waist circumference (WC) was measured at the narrowest level and that of hip at the maximal level over light clothing, using a tape measure. Waist hip ratio (WHR) was calculated as WC divided by hip circumference. To avoid subjective error, all measurements were taken by the same person. Blood pressure was measured by a qualified physician after a rest for 15 min.

Biochemical analysis

Blood samples (20 ml) were collected by venopuncture after 12-14 h fasting. Serum was separated by centrifugation at 3000 rpm for 15 min. A 100 μ l aliquot of serum was stored at -80° C until the measurement of serum PON1 activity. Biochemical measurements and HDL separation were carried out on the day of sampling. HDL-C and LDL-C were determined by direct enzymatic procedures; FBS,

total cholesterol (TC), and triglyceride (TG) were measured by enzymatic methods; and apo A-I and apolipoprotein B-100 (apo B-100) were measured by immunoturbidometric methods using an automated analyzer (Hitachi 902, Kyoto, Japan) and commercial kits (Diasys Diagnosis, Holzheim, Germany).

Serum PON1 activity

PON1 activity was measured using 2 mM paraoxon (diethyl p-nitrophenyl phosphate, Sigma chemical Co., St. Louis, MO, USA) as the substrate in 100 mM tris buffer (pH 8.0) containing 2 mM of $CaCl_2$. Aliquots of 40 μ l of serum were added to 500 μ l of substrate medium and the initial rate of hydrolysis (generation of p-nitrophenol) was monitored at 412 nm, using a continuously recording spectrophotometer (UV 3100, Shimadzu, Kyoto, Japan) over a period of 2 min. A PON1 activity of 1 unit/L was defined as 1 μ mol of p-nitrophenol formed per minute. The molar extinction coefficient of p-nitrophenol was taken at 17,000 M/cm. All assays were performed in duplicate at 25°C. The within-assay coefficient of variation (CV) for 10 separate measurements was 2.9% and between-assay CV for 10 separate measurements during 20 days was 3.6%.

HDL separation

For HDL separation (1.063 g/cm³< Q<1.21 g/cm³), polyallomer ultracentrifuge tubes with 8.9-ml capacity (Optiseal, Beckman/Coulter, Fullerton, CA, USA) were first taken with 5.9 ml of serum sample and the sample was overlaid with 3 ml of solution A [containing 0.195 mmol NaCl, 0.62 mmol NaOH, 0.01% (w/v) ethylenediaminetetraacetic acid disodium salt (EDTA-Na₂), d = 1.006 g/ml] in order to make discontinuous density gradient. The tubes were centrifuged in a Beckman Coulter Optima L-100 XP ultracentrifuge equipped with a type 90Ti fixed angle rotor at 60,000 rpm for 6 h at 16°C, acceleration: "5" and deceleration: "7". After centrifugation, the very low density lipoprotein (VLDL) fraction (the white layer at the supernatant) accompanied by 3 ml of the upper layer solution in tubes was removed and residual content of tubes was mixed with 3 ml of solution B (containing 24.8 g NaBr in 100 ml of solution A). The tubes were centrifuged at 60,000 rpm for 12 h at 16°C, acceleration:"9" and deceleration:"7". After centrifugation, the LDL fraction (appearing as a yellow-orange band at the supernatant) accompanied by 3 ml of the upper solution was removed. The residual content of tubes was mixed with 3 ml of solution C (containing 78.32 g of NaBr in 100 ml of solution A) and centrifuged for another 12 h at 16°C, acceleration:"9" and deceleration:"7". After centrifugation, the HDL fraction appearing as a yellow band at the supernatant was collected carefully and stored at -80°C until further analysis.[24] The protein concentration of the HDL fractions was determined according to a modified Lowry method.^[25]

HDL mean size determination

A zetasizer nano ZS instrument (Malvern, Worcestershire, UK) with a 532-nm green laser beam was used for the study. The scattered light was collected by detector at an angle of 173° using Non-Invasive Back-Scatter (NIBS) technology and directed to a correlator. The data were analyzed by zetasizer software (DTS, nano series, version 5.02, Malvern) and size information was reported as the Z-average by intensity.[26] All measurements were performed at 25°C in duplicate with automatic duration measurements. In order to determine the suitable solvent with the best reproducibility prior to commencing the measurements, six different solvents [including distilled water, normal saline, and 0.2 M phosphate buffer containing 0.1% (w/v) EDTA-Na, at three different pH values of 5.6, 7.4, and 8.3] were used as the HDL dispersant. Hourly measurements of size were made over a period of 5 h and the results were compared. Consequently, 0.2 M phosphate buffer at pH of 7.4 containing 0.1% (w/v) EDTA-Na, was selected as the most repeatable dispersant with the lowest aggregation index.

In order to measure HDL mean particle size by dynamic light scattering (DLS) methodology, isolated HDL samples were mixed gently with 1 ml of dispersant (at a final protein concentration of 250 µg/ml) and passed through a syringe filter (Millipore cellulose acetate membrane, 30 mm, 0.2 µm pore size) prior to injection into a disposable polystyrene cell (Malvern) in order to remove dust particles and were then subjected to size determination. Viscosity and refractive index (RI) of water as the dispersant were applied to standard operating protocol (SOP) prior to size determination. The accuracy of size measurements was examined using standard size nanoparticles [Gold Nanoparticles, 20 nm, 0.01% (w/v) aqueous solution, Nanocs Inc., New York, NY, USA] under the same experimental conditions and the results were matched with the diameter quoted by the manufacturer. The within-assay CV for 10 measurements was 1.2% and between-assay CV for 8 measurements was 3.1%. The detection limit of the assay for zetasizer instrument used in the present study was 0.3 nm to 10 µm.

HDL,/HDL, ratio

In order to calculate the ratio of HDL₂ to HDL₃, the accumulative percentage of particles with sizes larger than 8.2 nm was divided to the accumulative percentage of particles with sizes smaller than 8.2 nm.

Statistical analysis

Statistical analyses were performed using SPSS 16.0 (SPSS, Chicago, IL, USA). Before analyses, subjects were divided into three groups based on their serum HDL-C levels^[27] [group 1 (n = 49): HDL ≤ 39 mg/dl; group 2 (n = 46): 39>

HDL ≤49 mg/dl; group 3 (n = 45): HDL >49 mg/dl], and variables that were not normally distributed (i.e. TG, apo A-I and apo B-100) were subjected to natural log transformation. Quantitative parameters were compared between groups by analysis of variance (ANOVA) except for PON1 and HDL $_2$ /HDL $_3$ that were analyzed by a nonparametric Kruskal–Wallis test and data were expressed as means \pm SD. Correlations between variables with normal distribution were calculated by Pearson correlation coefficient analysis. Spearman's rho correlation coefficient analysis was used to calculate the correlation between variables that were not normally distributed (i.e. serum PON1 activity).

RESULTS

Clinical characteristic data of all subjects are summarized in Table 1. All subjects in the three groups were age matched. As seen in Table 1, anthropometric indices were significantly different among the three groups. The mean BMI value in group 3 subjects was lower compared with that in group 1 (P = 0.002). WHR in groups 2 and 3 was lower compared with that in group 1 (P < 0.05 and P = 0.001, respectively). There was a significant difference in the mean TG values among the three groups (groups 1 and 2, P < 0.001; groups 2 and 3, P = 0.002; and groups 1 and 3, P < 0.001), with group 3 having the lowest values. TC was higher in group 3 compared with those in groups 1 and 2 (P = 0.003 and P= 0.002, respectively). Apo A-I was also significantly higher in groups 2 and 3 in parallel with the increasing serum HDL-C levels. The serum PON1 activity level was lower in group 3 compared with those in groups 2 and 1 (P = 0.023and P = 0.004, respectively), whereas it was comparable in groups 1 and 2. PON1 activity adjusted for the HDL level (PON1/HDL-C) was also significantly lower in group 3 compared with those in groups 1 and 2 of our study (P < 0.001) and lower in group 2 compared with that in group 1 (P = 0.01). The mean HDL particle size and HDL₂/HDL₃ ratio was higher in the high HDL-C group compared with those in group 2 (P = 0.003 and P = 0.001, respectively) and group 1 (P < 0.001 and P < 0.001, respectively). The HDL₂/HDL₃ ratio also correlated with HDL-C levels in groups 1 and 2. There were no significant differences in blood pressure, FBS, LDL-C, and apo B-100 levels among the three groups.

Correlations between HDL-C, HDL mean particle size, $\mathrm{HDL}_2/\mathrm{HDL}_3$ ratio, and serum PON1 activity with anthropometric indices and clinical biomarkers are shown in Table 2. Serum HDL-C levels were positively correlated with TC, apo A-I, and LDL-C levels, and were negatively correlated with BMI, WHR, and TG (P < 0.01). HDL mean particle size and $\mathrm{HDL}_2/\mathrm{HDL}_3$ ratio were negatively correlated with BMI (P < 0.01), WHR, and serum TG levels (P < 0.05 and P < 0.01, respectively), and were positively correlated with serum HDL-C levels (P < 0.01). Results

Table 1: Clinical characteristics of the study participants									
Variable	Group 1	Group 2	Group 3	P value	P value	<i>P</i> value			
	(HDL ≤ 39)	$(39 < HDL \leq 49)$	(49 ≤ HDL)	(G1 and G2)	(G1 and G3)	(G2 and G3)			
Age (years)	31.35 ± 6.9	31.41 ± 8.4	35.16 ± 8.8	0.999	0.062	0.074			
BMI (kg/m²)	24.5 ± 3	23.2 ± 2.7	22.2 ± 3.2	0.125	0.002	0.281			
WHR	0.90 ± 0.05	0.87 ± 0.05	0.86 ± 0.05	0.031	0.001	0.576			
SP (mm Hg)	115.5 ± 8.9	115.9 ± 10.8	116.6 ± 11.4	0.986	0.860	0.930			
DP (mm Hg)	75.1 ± 5.7	77.2 ± 8.6	76.4 ± 11.4	0.317	0.632	0.850			
FBS (mg/dl)	85.3 ± 7.5	84 ± 7.3	83 ± 8.4	0.718	0.376	0.843			
TG (mg/dl)	2.28 ± 0.19	2.07 ± 0.17	1.94 ± 0.17	< 0.001	< 0.001	0.002			
TC (mg/dl)	176 ± 34	175 ± 33	203 ± 47	0.979	0.003	0.002			
HDL-C (mg/dl)	32.3 ± 4	43.5 ± 2.9	61.9 ± 7.2	< 0.001	< 0.001	< 0.001			
LDL-C (mg/dl)	102 ± 20	101 ± 14	109 ± 34	0.995	0.246	0.218			
apo A-I (g/L)	2.02 ± 0.04	2.06 ± 0.05	2.10 ± 0.08	0.010	< 0.001	0.005			
apo B-100 (g/L)	1.98 ± 0.10	1.93 ± 0.15	1.96 ± 0.14	0.213	0.776	0.588			
PON1 (U/L)	118.0 (39-162)	115 (41–162)	49 (38-68)	0.858	0.004	0.023			
PON1/HDL-C	3.6 (1.2-5.9)	2.7 (0.9-3.6)	0.8 (0.5-1.1)	0.01	< 0.001	0.001			
HDL mean size (nm)	8.54 ± 0.80	8.89 ± 0.86	9.52 ± 0.98	0.128	< 0.001	0.003			
HDL ₂ /HDL ₃ ratio	0.84 ± 0.69	1.07 ± 0.82	1.72 ± 1.2	0.031	< 0.001	0.001			

Table 2: Bivariate correlations between clinical biomarkers and HDL mean size, HDL₂/HDL₃ ratio, and PON1 activity in all subjects

Variable	Serum HDL-C	P value	HDLmean size	P value	HDL ₂ /HDL ₃ ratio	P value	PON1 activity	P value
Age (years)	0.198	0.19	0.51	0.546	0.022	0.797	-0.118	0.166
BMI (kg/m²)	-0.313	< 0.001	-0.262	0.002	-0.277	0.001	0.161	0.058
WHR	-0.279	0.001	-0.194	0.026	-0.186	0.032	0.178	0.042
SP (mm Hg)	0.064	0.464	-0.036	0.684	-0.043	0.622	-0.159	0.068
DP (mm Hg)	0.079	0.363	0.111	0.204	-0.071	0.416	-0.166	0.056
FBS (mg/dl)	-0.067	0.431	-0.185	0.029	-0.107	0.109	-0.049	0.570
TG (mg/dl)	-0.625	< 0.001	-0.204	0.015	-0.205	0.015	0.113	0.117
TC (mg/dl)	0.381	< 0.001	0.036	0.674	0.003	0.975	-0.116	0.173
HDL-C (mg/dl)	1	-	0.448	< 0.001	0.414	< 0.001	-0.247	0.003
LDL-C (mg/dl)	0.236	0.005	-0.038	0.652	-0.036	0.670	-0.051	0.554
apo A-I (g/L)	0.507	< 0.001	0.081	0.344	0.126	0.137	-0.215	0.011
apo B-100 (g/L)	0.014	0.872	-0.121	0.156	-0.082	0.336	-0.035	0.630

in Table 2 also indicate a significant negative correlation between serum PON1 activity and serum HDL-C levels (P < 0.01) and apo A-I (P < 0.05), and a significant positive correlation with WHR (P < 0.05).

Figure 1 shows individual correlations of serum PON1 activity with serum HDL-C level, HDL mean particle size, and HDL₂/HDL₃ ratio. As indicated, serum PON1 activity was negatively correlated with serum HDL-C level (r = -0.25, P = 0.003), HDL mean particle size (r = -0.22, P 0.008), and HDL₂/HDL₃ ratio (r = -0.23, P = 0.007). There was also a strong positive correlation between serum HDL-C level and HDL mean particle size (r = 0.45, P < 0.001) [Figure 2].

DISCUSSION

The present study describes the correlation between HDL mean particle size and serum PON1 activity and their relationship with anthropometric indices and lipid profile in healthy male subjects. We found a significant negative correlation between serum PON1 activity and HDL mean particle size in all subjects. Similar negative correlation was observed between serum PON1 activity and HDL₂/HDL₃ ratio. These findings are in accordance with Kontush's investigation which demonstrated that small HDL, particles posse higher PON1 activity than large HDL, particles.[17] Nishio's investigation on the susceptibility of HDL subfractions to copper-mediated oxidation indicated that HDL, is more susceptible to oxidation than HDL, ^[14] It is conceivable that this protective effect is mediated by PON1 activity being higher in small HDL₃ particles compared with large HDL, particles. There is increasing evidence suggesting that PON1 makes a central contribution to the antioxidant capacity of HDL. [9,28,29] However, Hasselwander's investigation has indicated that reduction in arylesterase activity of paraoxonase had no effect on the HDL ability to inhibit LDL oxidation in patients on regular hemodialysis.[30] Moreover, it has been demonstrated that HDL also exerts inhibitory effects on LDL oxidation, which are independent of paraoxonase activity.[31] Epidemiological

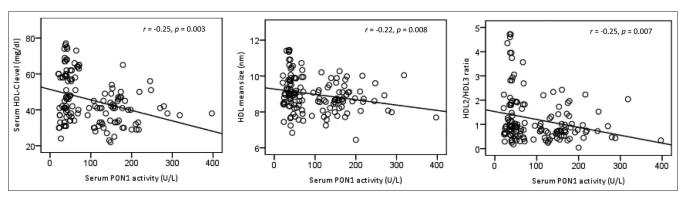


Figure 1: Correlations between serum PON1 activities with serum HDL-C levels, HDL mean particle size, and HDL-/HDL3 ratio

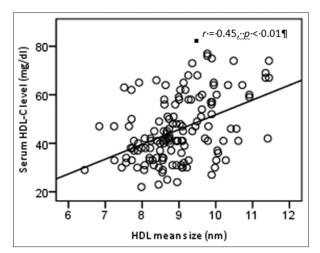


Figure 2: Correlations between HDL mean particle size and serum HDL-C levels

studies on the relationship between PON1 and CAD have reported conflicting results.[32] Several independent studies have indicated that the concentration and activity of PON1 are decreased in CAD patients.[20,33,33] Mackness et al. in a prospective study reported a reverse relationship between PON1 activity and CAD risk in men, [18] whereas in another prospective investigation on healthy middle-aged men, Troughton et al. did not find any effect of PON1 paraoxonase activity on coronary heart disease (CHD) outcomes.[19] Furthermore, van Himbergen et al. have reported that in a general population of Dutch women, increased PON1 paraoxonase activity was a risk factor for cardiovascular outcomes instead of being protective. [21] Our findings have indicated that serum PON1 activity in subjects with high HDL-C levels (group 3 of our study) is significantly decreased in comparison with subjects with medium and low HDL-C levels. This finding is in contradiction with a majority of investigations demonstrating that high PON1 levels and high PON1 activity contribute to increased levels of HDL-C.[17,35] In many examples of HDL deficiency, such as Tangier disease and fish eye disease, serum PON1 is significantly diminished, [36,37] but in other HDL deficiencies, this is not always the case. [38] Blatter et al. have reported a positive correlation between PON1 activity and HDL-C levels in control subjects and CAD patients.[39] They also demonstrated a significant decrease in PON1 concentration and its arylesterase and paraoxonase activities in CAD patients in comparison with control subjects. PON1 expression is partly controlled by its genetic variation.^[40] In addition, PON1 levels and activity could be modified by lifestyle determinants such as smoking, vitamin C and E consumption, and alcohol intake.[15] The contribution of PON1 genotypes to HDL-C levels was described by van Himbergen et al. who investigated on a population with familial hypercholesterolemia.[16] They described that 55 L, 192 R, -107C, and 907 G variants of the L55M, Q192R, -107C/T, and -907 G/C polymorphisms predicted increased HDL-C levels. They also indicated that like the PON1 genotypes, PON1 levels and paraoxonase, diazoxonase, and arylesterase activities also contributed to increased HDL-C levels. In our study, a strong positive correlation was found between serum HDL-C levels with HDL mean particle size and HDL₂/HDL₃ ratio. A similar direct relationship between HDL particle size and serum HDL-C Levels has been reported by others.[41,42] Decreased serum PON1 activity parallel to increased serum HDL-C levels observed in the present study may be a result of increased HDL particle size and the ratio of HDL₂/HDL₃, and consequently, decrease in the small dense HDL particles. In other words, changes in HDL subspecies distribution could explain the reduced activity of serum PON1. Shiavon et al. indicated that serum PON1 activity was decreased in chronic renal failure patients under maintenance hemodialysis.[43] They explained that reduction in PON1 activity was the consequence of the change in HDL subspecies distribution. They found that the PON1 enzyme activity on HDL was almost totally expressed by the HDL, subspecies fraction.[44]

Investigations have demonstrated a relationship between HDL particle size and CAD risk. The levels of small HDL₃b and HDL₃c (7.3-8.2 nm) were directly correlated with CAD severity, whereas the levels of intermediate HDL₃a

and HDL₂a (8.2-10 nm) and large HDL₂b (10-13 nm) were inversely correlated with CAD severity.[45] However, the protective role of HDL, has not been demonstrated in all studies.[21] Small dense HDL, has been shown to exert more powerful antioxidative, protective effects against LDL oxidation compared with the large HDL, subfraction.[17,46,47] As ox-LDL is the most atherogenic form of modified LDL, this protective role of HDL₃ particles is very important. Results of the anthropometric determinations in our study have indicated a reverse relationship between HDL mean particle size with BMI and WHR and a direct relationship between PON1 activity and these anthropometric indices. There are, however, a number of limitations of the present study. Because of the effects of female sex hormones such as estrogen and estradiol on lipid profile and PON1 activity and alterations of these hormones during menstrual cycles in women, in the present study we investigated only healthy male subjects; our results need to be verified in women. It is also recommended to determine the relationship between HDL mean particle size and PON1 activity in CAD patients in comparison with healthy subjects.

CONCLUSION

In conclusion, our results indicate that there is negative correlation between HDL mean particle size and serum PON1 activity in healthy Iranian male subjects. An inverse correlation also exists between HDL mean particle size and anthropometric indices, whereas plasma PON1 activity is directly correlated with them. Considering the complicated role of HDL in CAD prevention and its heterogeneous nature, further characterization of HDL subclasses, including particle size and its associated enzymes such as PON1, may be helpful for better understanding of CAD risks and prognosis and also may help to design therapeutic protocols toward beneficial modifications of HDL characteristics.

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REFERENCES

- Cooney M, Dudina A, De Bacquer D, Wilhelmsen L, Sans S, Menotti A, et al. HDL cholesterol protects against cardiovascular disease in both genders, at all ages and at all levels of risk. Atherosclerosis 2009;206:611-6.
- Maron DJ. The epidemiology of low levels of high-density lipoprotein cholesterol in patients with and without coronary artery disease. Am J Cardiol 2000;86:11L-14L.
- 3. Toth PP. Reverse cholesterol transport: High-density lipoprotein's

- magnificent mile. Curr Atheroscler Rep 2003;5:386-93.
- Cheung MC, Segrest JP, Albers JJ, Cone JT, Brouillette CG, Chung BH, et al. Characterization of high density lipoprotein subspecies: Structural studies by single vertical spin ultracentrifugation and immunoaffinity chromatography. J Lipid Res 1987;28:913-29.
- Perusse M, Pascot A, Despres JP, Couillard C, Lamarche B. A new method for HDL particle sizing by polyacrylamide gradient gel electrophoresis using whole plasma. J Lipid Res 2001;42:1331-4.
- Cho KH, Park SH, Park JE, Kim YO, Choi I, Kim JJ, et al. The function, composition, and particle size of high-density lipoprotein were severely impaired in an oliguric phase of hemorrhagic fever with renal syndrome patients. Clin Biochem 2008;41:56-64.
- 7. Kwiterovich PO Jr. Lipoprotein heterogeneity: Diagnostic and therapeutic implications. Am J Cardiol 2002;90:1i-10i.
- Murphy A, Chin-Dusting J, Sviridov D, Woollard K. The anti inflammatory effects of high density lipoproteins. Curr Med Chem 2009;16:667-75.
- Efrat M, Aviram M. Paraoxonase 1 Interactions with HDL, Antioxidants and Macrophages Regulate Atherogenesis A Protective Role for HDL Phospholipids. Adv Exp Med Biol 2010:153-66.
- Mackness B, Hine D, Liu Y, Mastorikou M, Mackness M. Paraoxonase-1 inhibits oxidised LDL-induced MCP-1 production by endothelial cells. Biochem Biophys Res Commun 2004;318:680-683.
- Deakin S, Leviev I, Gomaraschi M, Calabresi L, Franceschini G, James RW. Enzymatically active paraoxonase-1 is located at the external membrane of producing cells and released by a high affinity, saturable, desorption mechanism. J Biol Chem 2002;277:4301-8.
- James RW, Deakin SP. The importance of high-density lipoproteins for paraoxonase-1 secretion, stability, and activity. Free Radic Biol Med 2004;37:1986-94.
- 13. Wang M, Lang X, Cui S, Zou L, Cao J, Wang S, *et al.* Quantitative assessment of the influence of paraoxonase 1 activity and coronary heart disease risk. DNA Cell Biol 2012;31:975-82.
- 14. Fathy MM, Youssif OA, Ashour WM. The slow activity-associated genotype of microsomal epoxide hydroxylase-1 (m-EPHX-1) and low serum paraoxonase-1 (PON1) activity are associated with increased susceptibility to chronic obstructive pulmonary disease (COPD) in smokers. Comparative Clinical Pathology 2012:1-7.
- Lakshman R, Garige M, Gong M, Leckey L, Varatharajalu R, Zakhari S. Is alcohol beneficial or harmful for cardioprotection? Genes Nutr 2010;5:111-120.
- van Himbergen TM, Roest M, de Graaf J, Jansen EH, Hattori H, Kastelein JJ, et al. Indications that paraoxonase-1 contributes to plasma high density lipoprotein levels in familial hypercholesterolemia. J Lipid Res 2005;46:445-51.
- Kontush A, Chantepie S, Chapman MJ. Small, dense HDL particles exert potent protection of atherogenic LDL against oxidative stress. Arterioscler Thromb Vasc Biol 2003;23:1881-8.
- 18. Mackness B, Durrington P, McElduff P, Yarnell J, Azam N, Watt M, *et al.* Low paraoxonase activity predicts coronary events in the Caerphilly Prospective Study. Circulation 2003;107:2775-9.
- 19. Troughton JA, Woodside JV, Yarnell JW, Arveiler D, Amouyel P, Ferrières J, *et al.* Paraoxonase activity and coronary heart disease risk in healthy middle-aged males: The PRIME study. Atherosclerosis 2008;197:556-63.
- Mackness B, Davies GK, Turkie W, Lee E, Roberts DH, Hill E, et al. Paraoxonase status in coronary heart disease: Are activity and concentration more important than genotype? Arterioscler Thromb Vasc Biol 2001;21:1451-7.
- 21. van Himbergen TM, van der Schouw YT, Voorbij HA, van Tits LJ, Stalenhoef AF, Peeters PH, et al. Paraoxonase (PON1) and the risk for coronary heart disease and myocardial infarction in a general population of Dutch women. Atherosclerosis 2008;199:408-14.

- 22. Mehta RH, Califf RM, Garg J, White HD, Van de Werf F, Armstrong PW, et al. The impact of anthropomorphic indices on clinical outcomes in patients with acute ST-elevation myocardial infarction. Eur Heart J 2007;28:415-24.
- Beltowski J, Wojcicka G, Jamroz A. Leptin decreases plasma paraoxonase 1 (PON1) activity and induces oxidative stress: The possible novel mechanism for proatherogenic effect of chronic hyperleptinemia. Atherosclerosis 2003;170:21-9.
- Bronzert TJ, Brewer HB Jr. New micromethod for measuring cholesterol in plasma lipoprotein fractions. Clin Chem 1977;23:2089-98.
- Markwell MA, Haas SM, Bieber LL, Tolbert NE. A modification of the Lowry procedure to simplify protein determination in membrane and lipoprotein samples. Anal Biochem 1978;87:206-210.
- Rawle A. PCS in 30 minutes. Malvern, UK: Malvern Instruments Inc; 1995.
- Gharipour M, Baghaie AM, Boshtam M, Rabeie K. Prevalence of metabolic syndrome in An Iranian Adult population. ARYA Atherosclerosis 2010;1.
- Jayakumari N, Thejaseebai G. High prevalence of low serum paraoxonase-1 in subjects with coronary artery disease. J Clin Biochem Nutr 2009;45:278-84.
- Rosenblat M, Aviram M. Paraoxonases role in the prevention of cardiovascular diseases. Biofactors 2009;35:98-104.
- Hasselwander O, McEneny J, McMaster D, Fogarty DG, Nicholls DP, Maxwell AP, et al. HDL composition and HDL antioxidant capacity in patients on regular haemodialysis. Atherosclerosis 1999;143:125-33.
- 31. Chen X, Wu Y, Liu L, Su Y, Peng Y, Jiang L, *et al.* Relationship between high density lipoprotein antioxidant activity and carotid arterial intima-media thickness in patients with essential hypertension. Clin Exp Hypertens 2010;32:13-20.
- 32. Costa LG, Cole TB, Jarvik GP, Furlong CE. Functional genomic of the paraoxonase (PON1) polymorphisms: Effects on pesticide sensitivity, cardiovascular disease, and drug metabolism. Annu Rev Med 2003;54:371-92.
- 33. Jarvik GP, Rozek LS, Brophy VH, Hatsukami TS, Richter RJ, Schellenberg GD, *et al.* Paraoxonase (PON1) phenotype is a better predictor of vascular disease than is PON1(192) or PON1(55) genotype. Arterioscler Thromb Vasc Biol 2000;20:2441-7.
- Leviev I, Righetti A, James RW. Paraoxonase promoter polymorphism T(-107)C and relative paraoxonase deficiency as determinants of risk of coronary artery disease. J Mol Med (Berl) 2001;79:457-63.
- Saha N, Roy AC, Teo SH, Tay JS, Ratnam SS. Influence of serum paraoxonase polymorphism on serum lipids and apolipoproteins. Clin Genet 1991;40:277-82.
- 36. Mackness MI, Peuchant E, Dumon MF, Walker CH, Clerc M.

- Absence of "A"-esterase activity in the serum of a patient with Tangier disease. Clin Biochem 1989;22:475-8.
- 37. Mackness MI, Walker CH, Carlson LA. Low A-esterase activity in serum of patients with fish-eye disease. Clin Chem 1987;33:587-8.
- James RW, Blatter Garin MC, Calabresi L, Miccoli R, von Eckardstein A, Tilly-Kiesi M, et al. Modulated serum activities and concentrations of paraoxonase in high density lipoprotein deficiency states. Atherosclerosis 1998;139:77-82.
- Blatter Garin MC, Moren X, James RW. Paraoxonase-1 and serum concentrations of HDL-cholesterol and apoA-I. J Lipid Res 2006;47:515-20.
- Rojas-Garcia A, Solis-Heredia M, Pina-Guzman B, Vega L, Lopez-Carrillo L, Quintanilla-Vega B. Genetic polymorphisms and activity of PON1 in a Mexican population. Toxicol Appl Pharmacol 2005;205:282-9.
- Pascot A, Lemieux I, Prud'homme D, Tremblay A, Nadeau A, Couillard C, et al. Reduced HDL particle size as an additional feature of the atherogenic dyslipidemia of abdominal obesity. J Lipid Res 2001;42:2007-14.
- 42. Watanabe H, Soderlund S, Soro-Paavonen A, Hiukka A, Leinonen E, Alagona C, et al. Decreased high-density lipoprotein (HDL) particle size, prebeta, and large HDL subspecies concentration in Finnish low-HDL families: Relationship with intima-media thickness. Arterioscler Thromb Vasc Biol 2006;26:897-902.
- 43. Schiavon R, De Fanti E, Giavarina D, Biasioli S, Cavalcanti G, Guidi G. Serum paraoxonase activity is decreased in uremic patients. Clin Chim Acta 1996;247:71-80.
- 44. Schiavon R, Battaglia P, De Fanti E, Fasolin A, Biasioli S, Targa L, et al. HDL3-related decreased serum paraoxonase (PON) activity in uremic patients: Comparison with the PON1 allele polymorphism. Clin Chim Acta 2002;324:39-44.
- 45. Freedman DS, Otvos JD, Jeyarajah EJ, Barboriak JJ, Anderson AJ, Walker JA. Relation of lipoprotein subclasses as measured by proton nuclear magnetic resonance spectroscopy to coronary artery disease. Arterioscler Thromb Vasc Biol 1998;18:1046-53.
- Sakuma N, Yoshikawa M, Hibino T, Ohte N, Kamiya T, Kunimatsu M, et al. HDL3 exerts a more powerful antiperoxidative and protective effect against peroxidative modification of LDL than HDL2 does. J Nutr Sci Vitaminol (Tokyo) 2002;48:278-82.
- 47. Yoshikawa M, Sakuma N, Hibino T, Sato T, Fujinami T. HDL3 exerts more powerful anti-oxidative, protective effects against copper-catalyzed LDL oxidation than HDL2. Clin Biochem 1997;30:221-5.

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