Original Article

Evidence for an association of TP53 codon 72 polymorphism with sporadic colorectal cancer risk in Isfahan

Mehdi Nikbahkt Dastjerdi*, Mansoor Salehi**, Mohammad Reza Mohajeri***, Fereshteh Morsali****, Hamid Mirmohammad Sadeghi*****, Ebrahim Esfandiary*****

Abstract

BACKGROUND: A common polymorphism at codon 72 of TP53 gene has been associated with increased risk for many human cancers. We studied this TP53 polymorphism in colorectal adenocarcinomas in small population selected from Isfahan city.

METHODS: Samples: We undertook a case-control study on 180 controls and 180 paraffin block specimens of sporadic colorectal adenocarcinomas. PCR amplification of TP53 codon 72 polymorphism: TP53 codon 72 genotypes were detected by PCR using specific primer pairs for amplifying the Proline or the Arginine alleles.

STATISTICAL ANALYSES: The χ 2-test was used to assess the significance of any difference in the prevalence of TP53 codon 72 polymorphism between colorectal cancer patients and controls.

RESULTS: In control samples, the genotype distribution for TP53 polymorphism showed 28.3%, 48.9% and 22.8% for the Arginine/Arginine, Arginine/Proline and Proline/Proline genotypes, respectively. In the cancer group 40% of the cases were Arginine/Arginine, 42.2% were Arginine/Proline and 17.8% were Proline/Proline. A significant difference between cases and controls was found for the Arginine/Arginine genotype compared with (grouped) Arginine/Proline and Proline/Proline genotypes (Odds Ratio = 1.686 (1.085-2.620), P = 0.02).

CONCLUSIONS: TP53 codon 72 polymorphism may be a genetic predisposing factor for colorectal adenocarcinomas in Isfahan city.

KEYWORDS: Colorectal adenocarcinoma, TP53, Arginine, Proline, Polymorphism.

JRMS 2008; 13(6): 317-323

P53 is the most important tumor suppressor gene that is involved in many pathways such as apoptosis, cellular transcriptional regulation, and cell cycle control.^{1,2} The p53 protein has important role in cell cycle control, being involved in G1-phase arrest for DNA repairs or activation of the cell death machinery.³ The protein accumulates in the cytoplasm following DNA damage, and then translocates to the nucleus and activates gene transcription machinery for cell cycle arrest to allow repair of damaged DNA.⁴ Also p53 protein, in response to an excessive DNA damage, would activate programmed cell death pathway through transcriptional control of several genes.^{5,6} TP53, located on chromo-

JRMS/ November & December 2008; Vol 13, No 6.

^{*} Dept. of Anatomical Sciences, Medical school, Isfahan University of Medical Sciences.

^{**}Department of Genetics, Medical school, Isfahan University of Medical Sciences.

^{***}Lab. of Pathology, Isfahan, Iran.

^{****}Clinic of Dentistry, Isfahan, Iran.

^{*****}Dept. of Biotechnology in Pharmacy school, Isfahan University of Medical Sciences

^{******}Dept. of Anatomical Sciences, Medical school, Isfahan University of Medical Sciences.

Corresponding to: Dr Mehdi Nikbakht Dastjerdi, Dept. of Anatomical Sciences, Medical school, Isfahan University of Medical Sciences, Isfahan, Iran. E-mail: nikbakht@med.mui.ac.ir

some 17p13, is one of the most mutated genes affecting many types of human cancers.7,8 In addition to mutations, several polymorphisms in the wild-type TP53 gene locus have been detected, which could alter its function.9,10 Among the 14 polymorphisms identified in the TP53 gene, the most common in the general population associated with cancer development is the codon 72 Arg (Arginine) to Pro (Proline) substitution.¹¹ The TP53 Arg72Pro, located in exon 4 at codon 72 and involved in guanine to cytosine nucleotide exchange, leads to nonconservative change of an Arg to Pro. This polymorphism is located in a proline-rich region (residues 64-92) of TP53, homologous to the SH3 binding which is necessary for the protein to completely induce apoptosis.12 These two polymorphic forms of TP53 gene have different primary structure and electrophoretic migration properties¹¹ with some different biochemical and biological potentials including different binding to compartments of the transcriptional machinery and different ability to activation of transcription, but they do not differ in their property to bind DNA.13,14 Also the two polymorphic variants of TP53 have different potential to degradation by the HPV E6 protein.¹⁵ It is observed that the Pro variant involves in activation of transcription to a higher level compared to the Arg variant and it is better for inducing cell cycle arrest, while the Arg variant is able to induce apoptosis faster and more efficiently than the Pro variant.13

Colorectal cancer is one of the most common causes of cancer-related death in the world and it is the fourth commonest malignancy after lung, breast and prostate cancers.¹⁶ Many factors such as sex, age, diet and a variety of genetic factors influence the risk of developing colorectal cancer.^{17,18} There are a number of syndromes with Mendelian dominant inheritance in which there is a primary predisposition to benign or malignant tumors of colon. These syndromes are causes of only 2-6% of colorectal cancer cases. Most of the patients do not have Mendelian dominant inheritance, and many researchers suggest that other genetic factors might predispose the patients to this kind of cancer.¹⁹⁻²¹ Colorectal carcinogenesis is a complex multistage process that shows a high frequency of TP53 alterations and the large majority of these cancers are adenocarcinomas.^{18,22}

Because of functional differences between the two polymorphic variants of TP53, genotype at codon 72 may affect susceptibility to colorectal cancer development. TP53 codon 72 polymorphism has been associated with the risk of developing various human cancers such as lung,²³ esophageal,²⁴ cervical,²⁵ bladder,²⁶ breast,27-30 head and neck,31 pancreas,32 nasopharynx³³ and liver³⁴ cancers, although the results are still controversial. The role of codon 72 polymorphism of TP53 gene had been noted in the colorectal cancer patients in many popu-Argentina,35 lations including Taiwan,³⁶ Spain,³⁷ China,³⁸ Japan,³⁹⁻⁴¹ Turkey,⁴² Germany⁴³ France,⁴⁴ Sweden⁴⁵ and the USA.⁴⁶ It had been shown that the codon 72 polymorphism varies greatly in different ethnic populations⁴⁷ and this ethnic difference might have a significant effect on cancer risk in different ethnic populations. However, the role of the polymorphism in relation to colorectal cancer risk in the Iranian population has not been reported. This study explores a possible association between colorectal cancer and this polymorphism in a small population selected from Isfahan city.

Materials and methods

Study population and samples:

We performed a case-control study on 180 paraffin blocks of sporadic colorectal adenocarcinomas and 180 healthy controls, in order to examine possible associations between the Arg72Pro alleles and the risk of cancer. Incidental colorectal cancer cases (histologically confirmed) attending the Alzahra Hospital (Isfahan) over the period 2002-2006 made up the case group. Proximal tumors were defined as occurring in the cecum through to the transverse colon; tumors in the splenic flexure, descending and sigmoid colon, were defined as being distal. Other disorders of colorectal reThe TP53 codon 72 polymorphism and colorectal cancer

gion such as HNPCC, familial adenomatous polyposis, Inflammatory Bowel Disease (IBD), hamartoma, simultaneous occurrence of adenomas, previous or synchronous adenocarcinomas were excluded from this study. As control group, we used peripheral blood from 180 healthy age and sex matched persons. Controls were noncancerous persons who had already undergone colonoscopy.

DNA isolation from colorectal tissue and blood samples:

Genomic DNA from the tumors and blood samples was prepared using High pure PCR Template preparation DNA isolation kit (Roche, Germany) for tissue and whole blood, according to manufacturer's instructions.

PCR amplification of TP53 codon 72 polymorphism:

The TP53 codon 72 Pro allele was detected by PCR using the primer pair p53Pro+/ p53Pro-(p53Pro+: 5'-GCCAGAGGCTGCTCCCCC; and P53Pro-: 5'-CGTGCAAGTCACAGACTT) and the p53 codon 72 Arg allele by the primer pair p53Arg+/p53Arg- (p53Arg+:

5'-TCCCCCTTGCCGTCCCAA and p53Arg-: 5'-CTGGTGCAGGGGCCACGC).¹⁵ Between 100 to 300 nanograms DNA was used as template in a 25 μ l PCR reaction mixture containing 1.5 μ mol MgCl2, 1 U Taq polymerase (Sinagen) and 2 μ mol either of the primer pairs.

PCR cycling conditions were carried out with an initial denaturation step for 3 min at 94 °C, followed by 35 cycles of 30 s at 94 °C, 30 s at 60°C (for Arg) or 54 °C (for Pro) and 30 s at 72 °C. A final extension step was performed at 72 °C for 5 min. The PCR reaction was done separately for each of the two polymorphic variants. The amplified products were subjected to electrophoresis on 1% agarose gel in 1× TBE buffer and visualized on a transilluminator using ethidium bromide.

Statistical analyses:

The χ 2-test was used to assess the significance of any difference in the prevalence of TP53 codon 72 polymorphism between colorectal cancer patients and controls. The odds ratio and 95% CI (Confidence Intervals) were used as a measure of the strength of the association. Statistical significance level was set to $P \le 0.05$.

Results

This analysis included 180 adenocarcinomas and 180 cancer-free control subjects. The general and clinicopathological characteristics of the cases are shown in Table 1. The age of 180 patients (77 women and 103 men) ranged from 35 to 91 years (mean age of men 67.15 \pm 12.24 years, mean age of women 64.13 \pm 15.76 years).

Table 1. General and clinicopathological dataof patients with colorectal Cancer.

Factor	N (%)
Sex	
Male	57.2 (% 103)
Female	42.8 (%77)
Age	
≤ 59	67 (37.2 %)
≥ 60	62.8 (%113)
Localization	
Proximal	68.3 (% 123)
Distal	31.7(% 57)
Dukes stage	
A-B	23.9(% 43)
C-D	137 (76.1 %)
TNM staging	
Ι	25(% 45)
II	31.7(% 57)
III	36.1(% 65)
IV	13 (7.2 %)

To analyze the codon 72 polymorphism, we used a PCR-based assay that specifically amplifies either TP53 Pro or TP53 Arg allele and gives a PCR product by using specific primers for Pro allele (fig.1) and/or Arg allele (fig.2), respectively. Detection of TP53 codon 72 polymorphism by allele specific PCR was successfully conducted in all cases and controls. The distribution of the three different genotypes of codon 72 in exon 4 of TP53 in our cases and controls is shown in table 2. In control samples, the genotype distribution for p53 polymorphism showed 28.3%, 48.9% and 22.8% for the Arg/Arg, Arg/Pro and Pro/Pro genotypes, respectively. Allelic frequencies corresponded to 0.528 for the Arg allele and 0.472 for the Pro allele (table 3). In the cancer group, 40% of the cases were Arg/Arg, 42.2% were Arg/Pro and 17.8% were Pro/Pro (table 2). The corresponding frequencies in this group were 0.611 for the Arg allele and 0.389 for the Pro allele (table 3). A significant difference between cases and controls was found for

the Arg/Arg genotype compared with (grouped) Arg/Pro and Pro/Pro genotypes (Odds Ratio = 1.686 (1.085-2.620), P = 0.02). The Arg allele was found more often in patients than controls (Odds Ratio = 1.406 (1.064-1.891), P = 0.024).

Table 2. Distribution of TP53 codon72 polymorphism genotypes among colorectal cancer cases and controls in Isfahan

Construes	Cases(N	(=180)	Controls(1	N=180)	Odds Ratio
Genotype	Ν	%	Ν	%	(95% CI)
A/A	72	40%	51	28.3%	1.686*
A/P	76	42.2%	88	48.9%	
P/P	32	17.8%	41	22.8%	(1.085-2.620)

A/A: Arg/Arg genotype;

A/P: Arg/Pro genotype;

P/P: Pro/Pro genotype;

N: number

CI: Confidence Intervals

*: χ^2 test, P = 0.02 (Arg/Arg genotype compared with (grouped) Arg/Pro and Pro/Pro genotypes)

Table 3. Allelic frequencies of TP53 codon 72
among colorectal cancer cases and controls in
Isfahan

Allele	Patients	Controls	Odds Ratio (95% CI)
Arg	0.611	0.528	1.406*
Pro	0.389	0.472	(1.064-1.891)
CI: Confi	dence Interva	als	(

*: χ^2 test, P = 0.024

1 1 1 4 5 4 1 1

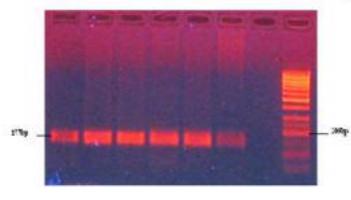


Figure 1. PCR amplification of the TP53 codon 72(electrophoresis in 1% agarose gel) in 6 colorectal adenocarcinoma specimens. lane 1-6: positive for Pro allele (177bp). lane7: negative control. lane8: DNA marker.

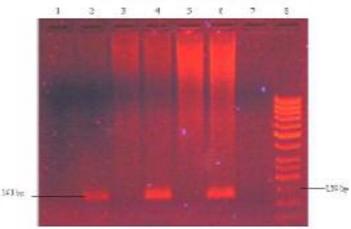


Figure 2. PCR amplification of the TP53 codon 72(electrophoresis in 1% agarose gel) in 6 colorectal adenocarcinoma specimens. lane 2, 4, 6 : positive for Arg allele (141bp). lane 1, 3, 5 : negative for Arg allele. lane7: negative control. lane 8: DNA marker.

Discussion

Molecular alterations have been associated with the development of colorectal cancer, including mutations at the TP53 tumor suppressor gene.²³ TP53 is polymorphic at amino acid 72 of the protein that it encodes, thus p53

protein may contain either an Arg or a Pro residue at this position. An association of the TP53 codon 72 polymorphism with several cancers susceptibilities has been reported.23-34 In particular, both Arg and Pro alleles have been shown to be associated with a high risk of malignancy. The role of the Arg/Pro polymorphism in colorectal cancer susceptibility has been examined in several studies,35-46 which have reported controversial results. We investigated the genotype frequencies of TP53 codon 72 in 180 sporadic colorectal adenocarcinomas and 180 healthy individuals from Isfahan and found a significant difference between cases and controls for the Arg/Arg genotype compared with (grouped) Arg/Pro and Pro/Pro genotypes (Odds Ratio = 1.686 (1.085-2.620), P = 0.02). The Arg allele was found more often in patients than in controls (Odds Ratio = 1.406 (1.064-1.891), P = 0.024). These findings are in agreement with the original study of Storey et al on cervical cancer.¹⁵ They showed that p53Arg72 protein is more susceptible to degradation by the HPV E6 proteins, and degradation of p53 protein by HPV E6 is correlated with increased risk for HPV-associated cancers. In this study we did not consider HPV infections in accordance to detection of p53 genotypes and it is an important issue for future studies. Our finding also seems to be consistent with the results reported by Perez et al³⁵ which support an appreciable association between the Arg allele and colorectal cancer. However, there are contradictory findings about the mechanisms which lead to the increase of the Arg allele in human cancers⁴⁸⁻⁵⁰ implicating that the involvement of TP53 polymorphism in human cancers demands further studies.

In contrast to our findings, other studies reported that the Pro allele had a significant effect on colorectal cancer risk,³⁶⁻³⁹ and some studies did not show a significant association of the polymorphism with colorectal cancer risk.⁴⁰⁻⁴⁶ The contradictory results about association of this polymorphism with colorectal cancer risk in different studies may be due to differences of allele frequencies between ethnic groups. It is well known that the distribution of TP53 codon 72 polymorphism varies in different geographic regions and ethnicities. According to the literature, general populations from Africa and Asia exhibit high frequencies of the Pro allele compared to the Arg one, while lower prevalences of Pro are found in populations of Latin America, the United States and Europe.^{47,51-53} In this study, the frequency for the Arg allele was higher than Pro one in the both cancer and control groups and therefore it is not consistent with the results of other studies on Asian populations.

It is already determined that this polymorphism acts as an intragenic modifier of mutant p53 behavior⁵⁴ and has an effect on the biological activity of p53, so the different genotypes of p53 my have different biochemical and biological potentials and may affect susceptibility to colorectal cancer development.

Other factors may interfere in colorectal cancer risk such as genetic heterogeneity in the pathogenesis of colorectal cancer, different environmental factors, and sample size limitations. It is also possible that TP53 codon 72 polymorphism could be in linkage disequilibrium with other putative etiological variants⁴⁵ which would likely differ across different ethnic populations.

The present study was not controlled for other potential predisposing factors, such as smoking or life-style habits. This is an important issue to be addressed in further studies in order to assess the role of TP53 polymorphism in this tissue.

In conclusion, the findings of the present study indicate that TP53 codon 72 polymorphism may be a genetic predisposing factor for colorectal adenocarcinomas and p53Arg72 protein may be correlated with possible increased risk of this kind of cancers in Isfahan.

Acknowledgement

This work was supported by Deputy for Research, Isfahan University of Medical Sciences (grant number 184002).

References

- 1. Dulic V, Kaufmann WK, Wilson SJ, Tlsty TD, Lees E, Harper JW et al. p53-dependent inhibition of cyclindependent kinase activities in human fibroblasts during radiation-induced G1 arrest. Cell 1994; 76(6):1013-1023.
- 2. Woods DB, Vousden KH. Regulation of p53 function. Exp Cell Res 2001; 264(1):56-66.
- 3. Sherr CJ. Principles of tumor suppression. Cell 2004; 116(2):235-246.
- 4. Kaelin WG, Jr. The p53 gene family. Oncogene 1999; 18(53):7701-7705.
- 5. Robles AI, Harris CC. p53-mediated apoptosis and genomic instability diseases. Acta Oncol 2001; 40(6):696-701.
- 6. Prives C, Hall PA. The p53 pathway. J Pathol 1999; 187(1):112-126.
- 7. Khan SA, Thomas HC, Toledano MB, Cox IJ, Taylor-Robinson SD. p53 Mutations in human cholangiocarcinoma: a review. Liver Int 2005; 25(4):704-716.
- 8. Borresen-Dale AL. TP53 and breast cancer. Hum Mutat 2003; 21(3):292-300.
- 9. Olivier M, Eeles R, Hollstein M, Khan MA, Harris CC, Hainaut P. The IARC TP53 database: new online mutation analysis and recommendations to users. Hum Mutat 2002; 19(6):607-614.
- **10.** Soussi T, Beroud C. Assessing TP53 status in human tumours to evaluate clinical outcome. Nat Rev Cancer 2001; 1(3):233-240.
- 11. Sreeja L, Syamala V, Raveendran PB, Santhi S, Madhavan J, Ankathil R. p °^rArg72Pro polymorphism predicts survival outcome in lung cancer patients in Indian population. Cancer Invest 2008; 26(1):41-46.
- **12.** Walker KK, Levine AJ. Identification of a novel p53 functional domain that is necessary for efficient growth suppression. Proc Natl Acad Sci U S A 1996; 93(26):15335-15340.
- **13.** Thomas M, Kalita A, Labrecque S, Pim D, Banks L, Matlashewski G. Two polymorphic variants of wild-type p53 differ biochemically and biologically. Mol Cell Biol 1999; 19(2):1092-1100.
- 14. Dumont P, Leu JI, Della PA, III, George DL, Murphy M. The codon 72 polymorphic variants of p53 have markedly different apoptotic potential. Nat Genet 2003; 33(3):357-365.
- **15.** Storey A, Thomas M, Kalita A, Harwood C, Gardiol D, Mantovani F et al. Role of a p53 polymorphism in the development of human papillomavirus-associated cancer. Nature 1998; 393(6682):229-234.
- 16. Parkin DM, Bray F, Ferlay J, Pisani P. Estimating the world cancer burden: Globocan 2000. Int J Cancer 2001; 94(2):153-156.
- 17. Rupnarain C, Dlamini Z, Naicker S, Bhoola K. Colon cancer: genomics and apoptotic events. Biol Chem 2004; 385(6):449-464.
- 18. Calvert PM, Frucht H. The genetics of colorectal cancer. Ann Intern Med 2002; 137(7):603-612.
- 19. Cannon-Albright LA, Skolnick MH, Bishop DT, Lee RG, Burt RW. Common inheritance of susceptibility to colonic adenomatous polyps and associated colorectal cancers. N Engl J Med 1988; 319(9):533-537.
- **20.** Houlston RS, Collins A, Slack J, Morton NE. Dominant genes for colorectal cancer are not rare. Ann Hum Genet 1992; 56(Pt 2):99-103.
- Johns LE, Houlston RS. A systematic review and meta-analysis of familial colorectal cancer risk. Am J Gastroenterol 2001; 96(10):2992-3003.
- 22. Gazelle GS, McMahon PM, Scholz FJ. Screening for colorectal cancer. Radiology 2000; 215(2):327-335.
- 23. Fan R, Wu MT, Miller D, Wain JC, Kelsey KT, Wiencke JK et al. The p53 codon 72 polymorphism and lung cancer risk. Cancer Epidemiol Biomarkers Prev 2000; 9(10):1037-1042.
- 24. Lee JM, Lee YC, Yang SY, Shi WL, Lee CJ, Luh SP et al. Genetic polymorphisms of p53 and GSTP1,but not NAT2,are associated with susceptibility to squamous-cell carcinoma of the esophagus. Int J Cancer 2000; 89(5):458-464.
- **25.** Zehbe I, Voglino G, Wilander E, Genta F, Tommasino M. Codon 72 polymorphism of p53 and its association with cervical cancer. Lancet 1999; 354(9174):218-219.
- **26.** Soulitzis N, Sourvinos G, Dokianakis DN, Spandidos DA. p53 codon 72 polymorphism and its association with bladder cancer. Cancer Lett 2002.14^r-1^vo:(^r)^v⁹;
- 27. Noma C, Miyoshi Y, Taguchi T, Tamaki Y, Noguchi S. Association of p53 genetic polymorphism (Arg72Pro) with estrogen receptor positive breast cancer risk in Japanese women. Cancer Lett 2004; 210(2):197-203.
- **28.** Kalemi TG, Lambropoulos AF, Gueorguiev M, Chrisafi S, Papazisis KT, Kotsis A. The association of p53 mutations and p53 codon 72, Her 2 codon 655 and MTHFR C677T polymorphisms with breast cancer in Northern Greece. Cancer Lett 2005; 222(1):57-65.
- **29.** Langerod A, Bukholm IR, Bregard A, Lonning PE, Andersen TI, Rognum TO et al. The TP53 codon 72 polymorphism may affect the function of TP53 mutations in breast carcinomas but not in colorectal carcinomas. Cancer Epidemiol Biomarkers Prev 2002; 11(12):1684-1688.
- **30.** Mahasneh AA, Abdel-Hafiz SS. Polymorphism of p53 gene in Jordanian population and possible associations with breast cancer and lung adenocarcinoma. Saudi Med J 2004; 25(11):1568-1573.

- **31.** Shen H, Zheng Y, Sturgis EM, Spitz MR, Wei Q. P53 codon 72 polymorphism and risk of squamous cell carcinoma of the head and neck: a case-control study. Cancer Lett 2002; 183(2):123-130.
- **32.** Dong M, Nio Y, Yamasawa K, Toga T, Yue L, Harada T. p53 alteration is not an independent prognostic indicator, but affects the efficacy of adjuvant chemotherapy in human pancreatic cancer. J Surg Oncol 2003; 82(2):111-120.
- **33.** Tsai MH, Lin CD, Hsieh YY, Chang FC, Tsai FJ, Chen WC et al. Prognostic significance of the proline form of p53 codon 72 polymorphism in nasopharyngeal carcinoma. Laryngoscope 2002; 112(1):116-119.
- 34. Yu MW, Yang SY, Chiu YH, Chiang YC, Liaw YF, Chen CJ. A p53 genetic polymorphism as a modulator of hepatocellular carcinoma risk in relation to chronic liver disease, familial tendency, and cigarette smoking in hepatitis B carriers. Hepatology 1999; 29(3):697-702.
- **35.** Perez LO, Abba MC, Dulout FN, Golijow CD. Evaluation of p53 codon 72 polymorphism in adenocarcinomas of the colon and rectum in La Plata, Argentina. World J Gastroenterol 2006; 12(9):1426-1. £79
- **36.** Lung FW, Lee TM, Shu BC, Chang FH. p53 codon 72 polymorphism and susceptibility malignancy of colorectal cancer in Taiwan. J Cancer Res Clin Oncol 2004; 130(12):728-732.
- **37.** Gemignani F, Moreno V, Landi S, Moullan N, Chabrier A, Gutierrez-Enriquez S et al. A TP53 polymorphism is associated with increased risk of colorectal cancer and with reduced levels of TP53 mRNA. Oncogene 2004; 23(10):1954-1956.
- **38.** Zhu ZZ, Wang AZ, Jia HR, Jin XX, He XL, Hou LF et al. Association of the TP53 codon 7 ^rpolymorphism with colorectal cancer in a Chinese population. Jpn J Clin Oncol 2007; 37(5):385-390.
- **39.** Kawajiri K, Nakachi K, Imai K, Watanabe J, Hayashi S. Germ line polymorphisms of p53 and CYP1A1 genes involved in human lung cancer. Carcinogenesis. 1. A9-1. A0: (1)15; 1997
- **40.** Murata M, Tagawa M, Kimura M, Kimura H, Watanabe S, Saisho H. Analysis of a germ line polymorphism of the p53 gene in lung cancer patients; discrete results with smoking history. Carcinogenesis 1996; 17(2):261-264.
- **41.** Hamajima N, Matsuo K, Suzuki T, Nakamura T, Matsuura A, Hatooka S et al. No associations of p73 G4C14-to-A4T14 at exon 2 and p53 Arg72Pro polymorphisms with the risk of digestive tract cancers in Japanese. Cancer Lett 2002; 181(1):81-85.
- **42.** Sayhan N, Yazici H, Budak M, Bitisik O, Dalay N. P53 codon 72 genotypes in colon cancer. Association with human papillomavirus infection. Res Commun Mol Pathol Pharmacol 2001; 109(1-2):25-34.
- **43.** Schneider-Stock R, Boltze C, Peters B, Szibor R, Landt O, Meyer F et al. Selective loss of codon 72 proline p53 and frequent mutational inactivation of the retained arginine allele in colorectal cancer. Neoplasia 2004; 6(5):529-535.
- 44. Olschwang S, Laurent-Puig P, Vassal A, Salmon RJ, Thomas G. Characterization of a frequent polymorphism in the coding sequence of the Tp53 gene in colonic cancer patients and a control population. Hum Genet 1991; 86(4):369-370.
- **45.** Sjalander A, Birgander R, Athlin L, Stenling R, Rutegard J, Beckman L et al. P53 germ line haplotypes associated with increased risk for colorectal cancer. Carcinogenesis 1995; 16(7):1461-1464.
- **46.** Koushik A, Tranah GJ, Ma J, Stampfer MJ, Sesso HD, Fuchs CS et al. p53 Arg72Pro polymorphism and risk of colorectal adenoma and cancer. Int J Cancer 2006; 11.1474-1477:(A)9
- **47.** Beckman G, Birgander R, Sjalander A, Saha N, Holmberg PA, Kivela A et al. Is p53 polymorphism maintained by natural selection? Hum Hered 1994; 44(5):266-270.
- **48.** Marin MC, Jost CA, Brooks LA, Irwin MS, O'Nions J, Tidy JA et al. A common polymorphism acts as an intragenic modifier of mutant p53 behaviour. Nat Genet 2000; 25(1):47-54.
- 49. Kaelin WG, Jr. The emerging p53 gene family. J Natl Cancer Inst 1999; 91(7):594-598.
- **50.** Monti P, Campomenosi P, Ciribilli Y, Iannone R, Aprile A, Inga A et al. Characterization of the p53 mutants ability to inhibit p73 beta transactivation using a yeast-based functional assay. Oncogene 2003; 22(34):5252-5260.
- **51.** Ojeda JM, Ampuero S, Rojas P, Prado R, Allende JE, Barton SA et al. p53 codon 72 polymorphism and risk of cervical cancer. Biol Res 2003; 36(2):279-283.
- 52. Zhu ZZ, Cong WM, Liu SF, Dong H, Zhu GS, Wu MC. Homozygosity for Pro of p53 Arg72Pro as a potential risk factor for hepatocellular carcinoma in Chinese population. World J Gastroenterol 2005; 11(2):289-292.
- **53.** Lu XM, Zhang YM, Lin RY, Liang XH, Zhang YL, Wang X et al. p53 polymorphism in human papillomavirusassociated Kazakh's esophageal cancer in Xinjiang, China. World J Gastroenterol 2004; 10(19):2775-2778.
- 54. Marin MC, Jost CA, Brooks LA, Irwin MS, O'Nions J, Tidy JA et al. A common polymorphism acts as an intragenic modifier of mutant p53 behaviour. Nat Genet 2000; 25(1):47-54.