Hypothalamic-pituitary-gonadal axis responses of the male rats to short and long time alternative magnetic fields (50 Hz) exposure

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Abstract

BACKGROUND: Electromagnetic fields are associated with production, transmission and use of electricity. In this study we have investigated the effects of short and long time alternative magnetic fields' (AMF, 50 Hz) exposure on the secretion of hypothalamic-pituitary-gonadal axis in the male rats.

METHODS: Forty eight Wistar male rats, same range of age and weight were divided into four groups and each group contained 12 rats. After one-week adaptation each group were exposed to AMF (0, 25, 50 and 100 $\mu$T respectively) for 17 days, 5 hours a day. In the second protocol the time of exposure extended to 34 days. After experiments rats' blood samples were removed from their blood samples and kept frozen for usage. The results were analyzed by one way- ANOVA statistical method (p < 0.05).

RESULTS: Chronic exposures (5h/day for 34 days) to AMFs had no effect on serum's testosterone and LH. But, AMF at 100 $\mu$T induced an increase of serum's FSH level in comparison with 25 $\mu$T, 50 $\mu$T and control groups. In contrast, sub-chronic AMFs (5 h/day for 17 days) induced a decrease of serum's testosterone in control group in comparison with 25, 50 and 100 $\mu$T groups. But these AMFs had no effect on serum's LH and FSH levels.

CONCLUSIONS: Increased level of FSH suggests damage to the seminiferous tubules. Our results suggest that AMFs probably causes dysfunction in gonadal axis at the hypothalamic-pituitary level in male rats in different protocols.

KEYWORDS: Sex Hormones, Male Rats, Magnetic Field.

Exposure of extremely low frequency (ELF, 50 and 60 Hz) electromagnetic fields (EMF) to human environment in the modern societies due to power stations, power cable transporting and many electrical appliances are very obvious, therefore the potential effects of ELF-EMF on human organs are very high and effective.1 ELF-EMF has possible adverse effects on reproductive and developmental outcomes which have studied extensively in animals in the past.2 Despite of the advantages of electricity there are some possible adverse effects too like being harmful for tissues, hormonal secretions, sperms process, development of proliferation and other biological cells.3-8 One area that has been studied by a number of researchers is the potential of EMFs to adversely affect reproduction.9 Studies have been, in most cases, evaluated the effects on the female, included effects on infertility, placental resorption, miscarriages, or embryo malformation.10 The possible effects of EMF exposure on male reproductive processes are few and experimental outcomes are quite different: reversible changes in spermatogenesis epithelium, aberrations in rat spermatozoa,
and no effects on human fertility.1 Forgacs et al indicated a presumably direct effect of whole body magnetic fields (MF) exposure on the chorionic gonadotropin-stimulated steroidogenic response of mice's leydig cells. MF exposure can make changes in neuroendocrine system's function in a mammalian laboratory model.12 Suppression of melatonin in the blood and pineal of rats and hamsters as a consequence of EMF exposure has been reported by several laboratories. Reduction in activity of N-acetyl transferase and the rate-limiting enzyme in the production of melatonin have been reported in a number of these studies too. Melatonin has been demonstrated to inhibit the hypothalamic-pituitary-gonadal (HPG) axis in certain species of mammals.12,13 There is also increasing concern that chronic or long-term exposure to low EMFs from many appliances, in the home or workplace, may cause adverse reproductive effects. Various researchers using mice as experimental models have attempted to elucidate the reproductive toxic effects of exposure to weak MFs and the results have been found to be rather contradictory.11 A significant increase in malformations was found after gestational exposure to a pulsed MF (20 KHz) in C3H mice. Al-Akhras et al have applied 25 \( \mu \text{T} \) sinusoidal MF for 90 days on adult male and female rats and have found some adverse effects on fertility of them.5

Also a lot of studies have been conducted on useful biological effects of EMF exposure on mammals, for example, Okano et al found that whole body exposure to static MF (SMF) at 10 and 25 mT for 2-9 weeks suppressed and delayed blood pressure (BP) elevation in young, stroke resistant, spontaneously hypertensive rats (SHR). They suggested in another study that SMF at 5 mT may suppress and delay BP elevation via the nitric oxide pathways and hormonal regulatory systems.15 But data about the effects on the male reproductive system are limited. There would seem to be a lack of agreement between scientists on the possible negative effects of EMFs on male reproductive parameters. Thus, there is an obvious need for additional information about the effects of exposure on HPG axis.

Some of previous studies have measured rat's mass, which are controversy.16-18 Some of the investigators reported increase,16 some reported decrease 17 and some reported no difference 18 between the weight of the control and electric or magnetic field treated rats, depending on the time of the exposure.

This study evaluated the possible effect of short and long time AMFs of 50 Hz on the secretion of testosterone, FSH and LH in the male rats, or the secretion of hypothalamic-pituitary-gonadal axis.

**Methods**

**Animal Maintenance**

Forty eight Wistar male rats, (obtained from Razi institute, Karaj, Iran) with same range of age (4-5 month old) and weight (215-230 g), were divided into four groups, each group contained 12 rats. After one-week for adaptation, each group was exposed to AMF (0, 25, 50 and 100 \( \mu \text{T} \) respectively), 5 hours daily for 17 days. In the second protocol the range of age was 2-3 month, weight was 63-70 g and the time of exposure was extended to 34 days. Experiments were performed using 4 alternative groups. To ignore several unwanted factors experiments were started at 9:00 A.M. Rats were housed; six per cage, with free access to tap water and fed laboratory chow (palletized commercial chow diet purchased from Pars Veterinary Food Company/Iran). They maintained at approximately 25°C and 55% humidity with a dark-light cycle of 12/12 hours. All procedures were performed in accordance with the guidelines of the University and National policies of animal for the care and use of laboratory animals, under an institutionally approved protocol.

**Experimental Procedure**

Exposure cages were made of transparent Kao-lin (resin) and each cage was equipped with top ventilation pores. Due to electrical safety conditions, water was withheld during the exposure period. The transparent cages and the
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open constructed coil systems allowed the animals to be easily viewed and to experience similar ambient conditions during both normal and exposure periods. In all experiments, the long axis of the cages was aligned with in direction of applied AMFs.

AMFs Generating Device

AMFs were designed and produced using homemade coil (Department of Medical Physics, Jundishapour Medical Sciences University of Ahwaz) which could generate static and pulsative MFs. The apparatus was electrical coil with up to 10000 turns and with nucleus rectangular cubical dimensions of $15 \times 15$ cm$^2$, made from pure iron in which the distance between poles could be varied and was adjustable. MFs were distinguished and measured using a gauss meter (Brockhause Messtechnic Company model 410) at different distances between and around the poles. Distances of cages were determined and rats were moved freely in these homogeneous fields. In both sides between the poles, the AMFs intensities were $25 \pm 1.25$, $50 \pm 2.5$ and $100 \pm 5$ mT and 6 rats were in each cage during exposure of AMF. Each field was determined using a gauss meter that we assured constancy of the field. The current could be activated through a switch which was housed in a locked box together with the transformer and resistor. The constancy of the fields could be controlled by the variation of the amperage which achieved by rheostat in the circuit of the system. The generated field did not cause any sound or any other manifestation that could be perceived by the rats or experimenters. Whole magnetic generator fixed to wooden framed boxes and the absence of significant vibrations was confirmed by acceleration measurements. Furthermore, the frame with coils was mechanically separated from the animals' cages. The coil's design (number of turns, cross-section of conductor and rated current) was optimized in order to reduce joule-heating losses to the level which did not affect the temperature of the exposure area. Control group's rats were held in a similar non-energized system in which the MF was equal to the background level ($< 0.05$ mT).

Collecting Blood Samples

Prior to exposure, animals were randomly assigned to one of the four exposure groups ($n = 12$). After 17 or 34 days of AMFs exposure, they anesthetized using ether and decapitated between 09:00 and 11:30 A.M. and their blood were immediately drawn into tubes without anticoagulant, allowed to clot, and then centrifuged to obtain the serum which was kept frozen under $-20^\circ$C until the time of usage. Hormones were measured using gamma counter equipment (Contron Company of Swiss) with RIA and IRMA methods.

Measurement of Testosterone, LH and FSH

Fifty $\mu$l of serum was used for the measurement of testosterone by radioimmunoassay (Biosource Testo-RIA-CT Kit). In this assay, the cross-reactivity with dihydrotestosterone is less than 1% and the minimal detectable concentration (M.D.C.) of testosterone was 0.44 ng/dl. The intra- and interassay variances (CV) were 4% and 8%, respectively. LH and FSH were measured using IRMA method (DSL Inc). In this assay, the cross-reactivity was non-detectable with other glycoprotein hormones (FSH, LH, TSH, HCG and $\beta$-hCG). M.D.C. of LH and FSH were 0.12 mIU/ml and 0.11 mIU/ml, respectively. The intra- and interassay variances of LH were 8% and 8%, and of FSH were 3% and 7%, respectively. Because rat's specific glycoprotein hormonal kit is expensive, human glycoprotein hormonal kits were used instead of animals' kits. Routine cross-reactivity test on rat's hypophysis ($n = 15$) also have been done for majority of experiments, which showed there are no differences between rat's LH and FSH (U.S.A.DRG International RIA catalog No.3732 and 3729, respectively) and human's kit in these circumstances.

Glycoprotein hormones' measurement must be done using rat's special kits. For this reason cross-reactivity test with human FSH and LH kits, was done on some of the rats which couldn't use rat's special kits and correlation coefficient ($r = 0.89$) obtained which showed there are much more similarity between humans' and rats' FSH and LH.
**Statistical Analysis**
Results are expressed as mean ± S.E.M. One way analysis of variance (ANOVA) which in some necessary cases, followed by, Tukey's test was used to determine the statistical significance of differences between means. A p value of less than 0.05 was taken as statistical significance.

**Results**
**Effect of The AMFs 50 Hz on Plasma Testosterone, FSH and LH**
There were statistically significant decreases in serum testosterone values in the groups that were exposed to AMFs for 17 days at 25, 50 and 100 µT in comparison with control group (Figure 1). These data showed that no changes in LH and FSH level were found at short time MFs exposure protocols in exposed or control group's animals. Testosterone hormone wasn’t changed significantly (p > 0.05) while exposing to AMFs for 34 days. These data showed that no changes in LH level were found at long time MFs exposure protocols were in exposed or control group's animals. FSH level of rats which exposed to 100 µT for 34 days increased significantly (p < 0.05) in comparison with those that exposed to 25 µT and 50 µT and control group's (Figure 2).

![Graph showing testosterone levels](image)

**Figure 1.** The level of testosterone concentration of the animals' group that exposed to 25, 50 and 100 µT AMFs, 5 hours daily for 17 days. Animals that exposed to 25 and 100 µT (p < 0.01 ) and 50 µT (p < 0.001 ) decreased significantly in comparisons with sham-exposed (n = 12). (Mean ± SE)

![Graph showing FSH levels](image)

**Figure 2.** The level of FSH concentration of the animals' group that exposed to 25, 50 and 100 µT AMFs, 5 hours daily for 34 days. Animals that exposed to 100 µT increased significantly (p < 0.05 ) in comparison with sham-exposed, 25 and 50 µT (n = 12). (Mean ± SE)

![Graph showing weight changes](image)

**Figure 3.** The weight of animals which have exposed to 25, 50 and 100 µT AMFs, 5 hours daily for 17 days. The sham and 100 µT groups have a significantly increase of weight (p < 0.01) in comparisons to the final stage of experiments but the weight of those animals which have been exposed to 25 and 50 µT have not changed significantly (n = 12). (Mean ± SE)
Effect of The AMFs 50 Hz on The Weight
Figures 3 and 4 have shown that animal's weight changed related to their age at the end of the exposures. Except those animals which exposed to 25 and 50 μT (17 days) AMFs (p < 0.05) (Figure 3).

Discussion
Effect of The AMFs 50 Hz on Plasma Testosterone, FSH and LH
We have studied possible effect of an alternative 50 Hz MFs with densities of 25, 50 and 100 μT exposure, 5 hours daily for 17 or 34 days, on hormonal changes of the control and exposed subjects.

Our data were shown that no changes of LH and FSH at any MFs protocols were found in exposed or control animals except FSH levels of rats which exposed to 100 μT (34 days) increased significantly in comparison with control group and 25 and 50 μT exposed. Also there was statistically significant decrease in serum testosterone values in those exposed to AMFs for 17 days, in comparison with control group.

However there are some controversial studies about the hypophysis-gonadal axis hormones which have been reported by different laboratories. Free et al (1981) have found alterations in the secretion pattern of FSH in rats exposed to an 80 KV/m electric field for 20 to 56 days. They have also observed a reduction in plasma's testosterone level after 120 days of electric field exposure.19 Al-Akhras et al (2006) reported that there were no significant effects on the serum's level of male follicle stimulating hormone (FSH) during the 18 weeks of exposure period. On the other hand, there was a significant increase in the serum's level of male luteinizing hormone (LH) after 18 weeks of exposing male rats to intensity magnetic field of 25 μT, while testosterone's level were significantly decreased only after 6 and 12 weeks of exposure. These results suggest that long term exposure to ELF could have adverse effects on mammalian fertility and reproduction.20 Al-Akhras (2008) reported that serum's level of luteinizing hormone (LH), follicle stimulating hormone (FSH), progesterone, and estrogen were measured before, after, and during the 18 weeks of exposing female rats to intensity magnetic field of 25 μT. Body and uterine weights were not affected by the field. The reduction in the levels of gonadotropins (FSH and LH) was significant after six weeks of exposure. FSH level was affected only on the sixth week of exposure while LH was still being affected after 12 and 18 weeks. Interestingly, no significant effects were found in 6 and 12 weeks after removing the field. The level of progesterone and estrogen was significantly decreased after 12 weeks of exposure.
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while no other effects on progesterone level was observed during exposure or after removing the field. The level of estrogen was also significantly reduced in 12 weeks after removing the field. These results suggest possible adverse effect on mammalian fertility and reproduction. The effects of ELF-MF on sex hormones were shown to be partly reversible. These are similar to the data that is being presented in this result. But Forgacs et al (1998) have demonstrated that exposure to sinusoidal, 50 Hz and 100μT MF increased the basal testosterone production in the primary 48 hours that mouse cultured leydig cell, whereas the steroidogenic capacity response to hCG remained unchanged.

These results suggest the probable causes of dysfunction in gonadal axis at the hypothalamic-pituitary level in male rats. So it can be concluded that AMFs were decreased the secretion of hypothalamic-pituitary-gonadal axis in male rats, and origin of disturbed testis in this protocol at least was related to hypothalamic-pituitary level.

Although our data show that there were no changes in serum's testosterone value in AMFs exposed groups for 34 days, in comparison with control group. This is similar to McGivern et al (1990) research that have reported pregnant Sprague-Dawley dams were exposed to a low-level, low-frequency pulsed EMF (15 Hz, 0.3 milli second duration, peak intensity 8 gauss) for 15 min twice a day from 15th day to 20th day of gestation (a period in development that is critical for sexual differentiation of the male rat brain). At the age of 120 days, field-exposed male offspring had normal circulating levels of the testosterone, LH and FSH hormones, as well as epididymal sperm counts. Also, Margonato et al (1993) who had done three years of investigation on the biological effects of high intensity electric field exposure to rats for up to 18% of their life span, but they did not find any differences on LH, FSH and testosterone between exposed animals and control group's animals. Kato et al (1994) reported that 6 weeks of nearly continuous exposure to circularly polarized 50 Hz magnetic fields did not change plasma's testosterone level in rats. This is similar to the data that is being presented in this paper. Margonato et al (1995) did not find any magnetic field-induced morphologic and histological changes in tested rats after prolonged exposure to a 50 Hz magnetic field at 5μT. Selmaoui et al (1997) in the same research direction reported that LH and FSH secretion was unaffected by the acute exposure to a 50 Hz linearly polarized magnetic fields of 10μT on 32 young men of 20-30 years old age. Zecca et al (1998) have chosen two groups of adult male Sprague-Dawley rats and exposed them to EMF of two different field strength combinations: 5μT-1KV/m and 100 μT-5KV/m for 8 months. They did not find any changes in serum's LH concentration. The latter finding is compatible with the absence of plasma's testosterone changes described by Kato et al (1994) in rats exposed to MFs of similar strength. Indeed, testosterone's release is mainly controlled by LH. These are similar to the data that is being presented in this study.

Effect of The AMFs 50 Hz on The Weight

Weight's changes experiments show different patterns according to various protocols. Our data about the weight's changes in those exposed for 34 days 5 hours a day, is the same as the report of Hilton and Phillips. There were no differences between effects of MFs on weight of the control group and electric or magnetic field treated rats. In this, protocol we have not seen any significant changes in the weight due to AMFs exposure which can be speculated according to no change in the testosterone concentration.

In conclusion the results of this study show the effects of AMFs on the secretion of hypo-
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thalamic-pituitary-gonadal axis in male rats. Present results show that there was statistically significant decrease in HPG axis at short duration (17 days). In our views the effects of EMFs on the biological systems depend on the intensity and time of exposure. We have reported that short duration (17 days) of exposing to SMFs have no effects on hypothalamic-pituitary-gonadal and thyroidal hormones. In contrast we have also reported that long term (34 days) and increased daily time exposure to SMFs can alter hypothalamic-pituitary-gonadal and thyroidal hormones.

Conclusions
Therefore we can conclude that MFs have widespread biological effects especially on sex organ and CNS. The melatonin can be involved to affect on dysfunction in gonadal axis at the hypothalamic-pituitary level in rats exposed to 50 Hz electromagnetic fields. The mechanism of action is unknown.

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Conflict of Interests
Authors have no conflict of interests regarding this paper.

Authors' Contributions
AA and HFM designed the study, collected and analyzed the data and wrote the manuscript. MJTB operated and controlled magnetic fields, HS endocrinology consultant and MB helped in statistics analysis.

All authors have read and approved the content of manuscript.

References